Synthesis of the Spiroiminal Moiety and Approaches to the Synthesis of Marineosins A and B

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Supporting Information

ABSTRACT: A short and efficient synthesis of model spiroiminals that have the same stereochemistry as marineosins A and B, but different conformations, was carried out in six or seven steps from 6-methyltetrahydropyran-2-one. These spiroiminals were also prepared biomimetically by reduction of an enol ether. A more highly substituted spiroiminal with the same stereochemistry and conformation as marineosin A was prepared in 11 steps from parasorbic acid. A macrocyclic pyrrole lactone was prepared stereospecifically in 10 steps. A five-step sequence converted the lactone to a late hemiiminal intermediate that has resisted the methylation and spiroiminal formation that would lead to marineosin A.



INTRODUCTION

In 2008 Fenical and co-workers isolated the cytotoxic spiroiminals marineosins A (1) and B (2) from a marinederived *Streptomyces*-related actinomycete (see Scheme 1).¹

Scheme 1. Structures and Biosynthesis of Marineosins A and B



The structures were determined by analysis of the NMR spectra with the stereochemistry assigned by interpretation of the NOESY spectra. Marineosins A and B differ in stereochemistry at both C-7 and C-8. MMX calculations and examinations of models suggest that the marineosin isomers at the spiroiminal center (C-8) are much less stable than the two isolated isomers because of steric interactions between the methoxy group and the adjacent pyrrole in the macrocycle. The major isomer marineosin A (1) inhibited human colon carcinoma HCT-116 with an IC₅₀ of 0.5 μ M, and testing in the NCI 60 cell line panel showed considerable selectivity against melanoma and leukemia cell lines. In contrast, marineosin B (2) showed considerably weaker cytotoxicity against human colon carcinoma HCT-116 with an IC_{50} of 46 $\mu \rm M.$

Marineosins A (1) and B (2) are novel members of the prodigiosin family of bacterial pigments that appear to be derived from an undecylprodiginine (undecylprodigiosin).² There are many examples of both spiroaminals and iminals, but the spiro-tetrahydropyran-dihydropyrrole (spiroiminal) moiety of the marineosins appears to be unprecedented. Fenical proposed that the biosynthesis of marineosins A and B involves an inverse electron demand intramolecular Diels–Alder reaction with a side chain enone as the diene to give a dihydropyran that undergoes a four-electron reduction to give the marineosins.¹ Lindsley³ and Haran⁴ established that this Diels–Alder reaction could not be achieved in the laboratory, suggesting that it is not the biosynthetic route.

Reynolds and Salem sequenced the gene cluster responsible for the biosynthesis of marineosins A and B in *Streptomyces* CNQ-617.⁵ The enzyme MarG, a RedG homologue from the *mar* gene cluster, oxidizes hydroxyundecylprodigiosine **3** at the asterisked carbon. Subsequent macrocyclization and spiroiminal formation affords dehydromarineosin A (4). The enzyme MarA, a putative dehydrogenase/reductase, catalyzes the reduction of **4** to afford marineosin A (1).

In 2010, we communicated the synthesis of spiroiminal models 18-21.⁶ Shi recently reported a very different approach to spiroiminals 18b-21b,⁷ and Lindsley prepared analogous spiroiminals lacking the methyl group.⁸ Lindsley also reported the synthesis of the functionalized macrocyclic pyrrole core of marineosin A.⁹

We report here the full details of our spiroiminal model studies, including those with a fully substituted tetrahydropyran

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ring with the marineosin A stereochemistry and conformation. We also describe an approach to marineosin A that leads to a fully functionalized macrocyclic core lacking the spiroiminal ring. Our synthetic plan is shown in retrosynthetic form in Scheme 2. The synthesis of 1 and 2 will be completed by acid-

Scheme 2. Retrosynthesis of Marineosins A and B



catalyzed spiroiminal formation of methoxy iminal **5** and pyrrole deprotection. Hydrogenolysis of isoxazoline **6** over Raney nickel should give a hemi-iminal that will be methylated to give **5**. Addition of a vinyl anion to lactone 7 and protection of the alcohol will give a vinyl ketone that will react with a protected pyrrole nitrile oxide to give isoxazoline **6**, most likely as a mixture of diastereomers that will both be elaborated to both marineosins A or B. Ring-closing metathesis of diene **8** and hydrogenation will construct the macrocycle of **7**. Conjugate addition of an allyl group to **9** should occur by axial attack from the face opposite the methyl group. Equilibration should give the desired stereoisomer of **8** with equatorial allyl and hexenylpyrrole groups. Pyrrole lactone **9** will be prepared by a Suzuki coupling of pyrrole boronic acid **10** and iodolactone **11**.

RESULTS AND DISCUSSION

We started with a model study to prepare phenyl spiroiminals 18a-21a for two reasons (see Scheme 3). The unprecedented spiroiminal moiety is the most intriguing, but also most challenging, moiety of the marineosins (1 and 2). The phenyl group is more stable than the pyrrole group¹⁰ and will allow us to first address the spiroiminal moiety without worrying about the instability of the pyrrole.

Treatment of readily available model lactone 12 with vinylmagnesium bromide afforded the known hydroxy ketone 13a in 85% yield (see Scheme 3).¹¹ Benzaldehyde oxime was



treated with NCS at room temperature to provide the chloro oxime, which was cooled to -78 °C and treated with Et₃N to generate benzonitrile N-oxide, which was treated with enone 13a to provide the hemiketal form of the isoxazoline in 75% vield as a 1:1 mixture of two diastereomers (see eq 1). Torsell reported in 1983 that hydrogenolysis of the isoxazolinyl methyl ketone analogous to 14a gave a hemi-iminal.¹² Vinyl ketone 13a exists preferentially in the open form because conjugation energy (~3 kcal/mol) is lost on cyclization to the hemiketal. However, once the isoxazoline is formed, the hemiketal dominates (>95%) in the equilibrium between the hemiketal and the corresponding saturated hydroxy ketone. Therefore it is not surprising that hydrogenolysis of the isoxazoline hemiketal formed from 13a over Raney nickel to reduce the isoxazoline failed to form a hemi-iminal by cyclization of the imine to the ketone. A variety of other reduction approaches were also unsuccessful.13



To prevent hemiketal formation, we protected the hydroxy group of 13a with TESCl, Et₃N, and DMAP to give 13b in 96% yield. Reaction of benzaldehyde oxime, NCS, and Et₃N at 25 °C generated benzonitrile *N*-oxide, which was cooled to -78 °C and treated with 13b to give the [3 + 2]-cycloadduct isoxazoline ketone 14a in 78% yield as a 1:1 mixture of diastereomers. As expected, treatment of isoxazoline ketone 14a with Raney Ni 2800 and H₂ in MeOH afforded the desired hemi-iminal 15a as a mixture of diastereomers. Unfortunately, attempted deprotection of the silyl ether under a variety of acidic conditions resulted in decomposition, rather than formation of the desired hydroxy spiroiminal. Hemi-iminal 15a even decomposed in CDCl₃ (containing adventitious HCl) in 10 h. We therefore treated 15a with sodium hydride and

methyl iodide to afford methyl ether iminal 16a in 58% yield from 14a.

The conversion of 16a to spiroiminals 18a-21a was explored under a variety of acidic conditions (see Scheme 4).





Treatment of 16a with 2% TFA in chloroform or PPTS in MeOH resulted in decomposition. Treatment of 16a with HF-Pyr and pyridine in THF gave the desired spiroiminals 18a-21a in only 15% yield and the undesired methoxypyrrole 17a in ~50% yield. Finally, we found that treatment of 16a with 2 M HCl in 1:1 THF/CH₃CN afforded three of the four desired spiroiminal diastereomers, 18a (46%), 19a (13%), and 21a (12%), and only 8% of the undesired methoxypyrrole 17a. Over a 2-3 week period in CDCl₂ (containing adventitious HCl), solutions of either pure 19a or 21a equilibrated to an identical 3:1 mixture of 19a and 21a. Therefore, these two spiroiminals differ only at the iminal center C-5 and have the identical relative stereochemistry at C-4 and C-7. Under the same conditions, the major isomer 18a equilibrated to give a 19:1 mixture of 18a and 20a. Therefore, these two compounds also differ only at the iminal center C-5. Attempts to accelerate the equilibration of 18a-21a by addition of 2% TFA to CDCl₃ resulted in the formation of methoxypyrrole 17a.

The structure of **21a** was established by an NOE between the CHOMe proton H-4 and the CHMe proton H-7 as shown in Scheme 4. In the other three isomers these two protons are too far apart for an NOE to be observed. The structure of **19a** follows from the structure of **21a** because these compounds differ only in the stereochemistry at the spiroiminal center. The structure of **19a** was confirmed by NOEs between the CHOMe proton H-4 and the adjacent tetrahydropyran methylene group as shown in Scheme 4. The CHMe proton H-7 in **18a** (δ 4.41) and **19a** (δ 4.45) is deshielded by the axial nitrogen and absorbs much further downfield than the CHMe proton H-7 in **20a** (δ 3.78) and **21a** (δ 3.79) with an axial carbon.¹⁴ The major isomer **18a** has no NOEs as expected between the protons on the tetrahydropyran ring and those on the dihydropyrrole ring.

The presence of the imine double bond makes the formation of spiroiminals from **16a** quite different from that of spiroketals

and spiroaminals.¹⁵ Desilylation should occur easily to give the alcohol. Protonation of the resulting alcohol on the iminal methoxy group and loss of MeOH would give a stabilized allylic type cation $C^+-N=C$ that could cyclize to form the spiroiminal, but the nitrogen lone pair cannot stabilize the cation by resonance because the five-membered ring precludes a linear cumulene $C=N^+=C$. Formation and equilibration of the spiroiminals could also occur by initial isomerization of the imine to an enamine or by protonation on the imine nitrogen and ring opening of the dihydropyrrole ring to give an oxycarbenium ion.

Having developed a sequence to make phenyl-substituted spiroiminals 18a-21a, we turned our attention to making pyrrole-substituted spiroiminals 18b-21b, which have the same spiroiminal moieties as marineosins A and B (1 and 2). Treatment of 13b with 2-pyrrolecarboxaldehyde oxime in THF with NCS and Et₃N in THF at -78 °C provided isoxazoline 14b in 62% yield. Hydrogenolysis of 14b over Raney nickel provided hemi-iminal 15b, but selective methylation of the hydroxyl groups without methylation of the pyrrole could not be achieved. The N-Me dimethyl ether 16d was obtained in 41% yield from 14b with NaH and MeI. Other methylation conditions were investigated unsuccessfully. Treatment of N-Me dimethyl ether 16d with 2 M HCl afforded N-Me pyrrole spiroiminals 18d, 19d, and 21d in 65% yield as a mixture of three diastereomers whose structures were assigned by analogy to 18a, 19a, and 21a.

We therefore needed to protect the pyrrole *N*-H to prevent *N*-methylation. Oxidation of *N*-Boc-2-pyrrolecarboxaldehyde oxime with $PhI(OAc)_2^{16}$ generated *N*-Boc-2-pyrrole-2-carbonitrile *N*-oxide, which added to enone **13b** to produce the desired isoxazoline **14**, Ar = *N*-Boc-pyrrol-2-yl, in 55% yield. However, treatment of the *N*-Boc pyrrole isoxazoline with Raney Ni and hydrogen not only cleaved the N–O bond of the isoxazoline but also hydrogenated the *N*-Boc pyrrole. The strong electron-withdrawing group on the pyrrole nitrogen reduces the aromaticity of the pyrrole ring making the pyrrole susceptible to hydrogenation.¹⁷

A SEM-protected pyrrole should be compatible with the hydrogenation step.^{18,19} N-SEM-pyrrole-2-carboxaldehyde¹⁸ was easily converted to the oxime with hydroxylamine hydrochloride and sodium acetate in aqueous MeOH. However, the oxidation conditions (NCS or iodobenzene diacetate) that were successful with other oximes gave 14c in <30% yield. Fortunately, reaction of N-SEM-pyrrole-2-carboxaldehyde oxime with 5% aqueous NaOCl²⁰ in CH₂Cl₂ at 25 °C generated the nitrile N-oxide that reacted with enone 13b to give isoxazoline 14c in 73% yield. Hydrogenolysis over Raney Ni and methylation both now proceeded uneventfully to give 16c in 42% yield from 14c. Treatment of 16c with 2 M aqueous hydrochloric acid in 1:3 THF/CH₃CN hydrolyzed the triethylsilyl ether and effected loss of methanol and cyclization to give SEM-protected spiroiminal 18c (34%), an inseparable 3:2 equilibrium mixture of SEM-protected spiroiminals 19c and **21c** (34%), and <2% of methoxypyrrole **17c**. The ¹H and ¹³C NMR spectra of these spiroiminals in the aliphatic region spiroiminals are virtually identical to those of 18a-21a, and their stereochemistry was assigned accordingly.

The initial model study was completed by deprotection of **18c** with TBAF and molecular sieves in THF at 60 $^{\circ}$ C for 3 h to provide spiroiminal **18b** in 54% yield (see Scheme 5). Similarly, deprotection of the 3:2 mixture of **19c** and **21c** afforded a 7:3 mixture of **19b** and **21b** in 56% yield. The





stereochemistry was again assigned from the NMR spectra, which are very similar to those of 18a-21a.

Salem and Reynolds's biosynthetic studies established that the last step of the biosynthesis of marineosin A (1) is the reduction of the enol ether of 4 to give 1.⁵ We wanted to explore this approach for the preparation of model spiroiminals 18–21. Berner and co-workers reported that oxidation of 3methoxypyrrole 22 with MnO₂ in acetone afforded 23 in 73% yield (see Scheme 6).²¹ We thought that the analogous





oxidation of 3-methoxypyrrole 17a would give 24 and 25, with the oxidized intermediate trapped intramolecularly by the hydroxy group in the side chain rather than intermolecularly by water. 3-Methoxy-5-phenylpyrrole 17a was a minor byproduct (8%) in the cyclization of 16a to 18a–21a with HCl in THF/ CH₃CN but was formed in ~50% yield on treatment of 16a with HF·pyr and pyr. Further optimization led to the formation of 17a in 65% yield by treatment of 16a with HF in THF. As expected, treatment of 17a with MnO₂ in acetone gave a mixture of easily separated spiroiminals 24 (14%) and 25 (35%), whose stereochemistry was assigned by hydrogenation to 18a–21a.

Hydrogenation of the major isomer 25 over Pd/C afforded a mixture of over-reduced spiroaminal diastereomers in \sim 70%

yield in which both the imine and enol ether double bonds had been hydrogenated. Hydrogenation of 25 over Raney Ni gave a mixture of the previously prepared spiroiminals 20a (15%) and 21a (15%) in addition to 3-methoxy-5-phenylpyrrole 17a (45%) resulting from hydrogenolysis. The best results were obtained by hydrogenation of 25 over 5% Pd/BaSO4, which afforded the desired spiroiminals 20a (40%) and 21a (20%) and only 10% of 3-methoxypyrrole 17a. Similarly, hydrogenation of the minor isomer 24 over 5% Pd/BaSO₄ provided 18a (35%), 19a (26%), and 17a (10%). We were pleased to find that spiroiminal 20a, which has the same stereochemistry as marineosin A (1) was obtained in 40% yield by hydrogenation of 25. This isomer was formed in trace amounts by the HCl-catalyzed spiroiminal formation from 16a and was formed in only $\sim 5\%$ yield during the equilibration of 18a in CDCl₂ over 2 weeks.

3-Methoxy-5-phenylpyrrole 17a was obtained in 65% yield by treating 16a with HF in THF. Unfortunately treatment of *N*-SEM-pyrrole dimethyl ether 16c with HF·Pyr or HF in THF and a variety of other acidic conditions gave a complex mixture rather than the desired 2,2'-bi-1*H*-pyrrole 17c, so this approach cannot be used to prepare 18c-21c.²²

Marineosin A (1) and model spiroiminal 20a have the same stereochemistry but very different conformations. As expected 20a adopts the conformation with equatorial nitrogen and methyl groups, whereas the macrocyclic ring of 1 locks the conformation of the tetrahydropyran ring so that both the nitrogen and methyl groups are axial. We therefore next turned to the preparation of a more highly substituted model lactone that would lead to a spiroiminal with the same conformation as marineosin A.

Both the second model study and the marineosin A synthesis start with (\pm) -parasorbic acid (26),²³ but enantiomerically pure **26** can be easily prepared once the sequence is worked out (see Scheme 7).²⁴ Parasorbic acid (**26**) was treated with iodine and

Scheme 7. Preparation of Vinyl Ketone 33



pyridine to afford iodolactone **11** in 74% yield. Suzuki coupling of iodolactone **11** with phenylboronic acid and 10 mol % $Pd(PPh_3)_4$ afforded phenyl lactone **27** in 47% yield. The yield was increased to 85% by using Buchwald's $Pd(OAc)_2$, SPhos, and *n*-butanol conditions.²⁵

All attempts to achieve 1,4-addition of an allylcuprate to 27 were unsuccessful as had previously been noted for a related unsaturated lactone by Waldmann and co-workers.²⁶ Trauner and co-workers reported that treatment of a variety of enones with allyltributyltin and Tf₂O afforded the corresponding vinyl triflates that can be used for intramolecular Heck reactions.²⁷ Unsaturated lactone 27 reacted similarly with allyltributyltin, di*tert*-butyl peroxide, and Tf₂O to afford the vinyl triflate in 78% yield, which underwent Stille coupling with tributylvinyltin and Pd(PPh₃)₄ to afford 30 in 83% yield. Cycloaddition of 30 with *N*-SEM-pyrrolecarboxaldehyde oxime and NaOCl occurred selectively as desired on the vinyl group of 30 in 58% yield. However, we were not able to hydrolyze the dihydropyran enol ether to give the required hydroxy ketone precursor for the Raney nickel hydrogenation step.

Fortunately, treatment of unsaturated lactone 27 with allylmagnesium bromide, ZnBr2, and TMSCl as described by Waldmann afforded a 1:3 to 1:1 mixture of the desired conjugate addition products 28a and 29a in 63-72% yield. We expected that conjugate addition of the allyl group would occur by axial attack from the face opposite the methyl group. Treatment of the mixture with DMAP in CDCl₃ provided a 6:1 equilibrium mixture favoring the desired trans isomer 28a. The equilibration of 28a and 29a establishes that they differ only in the stereochemistry at the phenyl-substituted carbon. The vicinal coupling constants between the methine hydrogens establish that the phenyl and allyl groups are trans in 28a (J =9.8 Hz) and *cis* in **29a** (J = 5.6 Hz). The stereochemistry of **28a** was confirmed by a strong NOE between the CHPh proton at δ 3.46 and CHMe at δ 4.66–4.56, which indicates that these two protons are mast protons in the expected boat conformer²² of 28a. The allyl double bond is needed for the ring-closing metathesis in the synthesis, but not for the model study, so the 6:1 mixture of 28a and 29a was hydrogenated over Raney nickel to afford a 6:1 mixture of trisubstituted lactones 28b and 29b

Attempted addition of vinylmagnesium bromide to the 6:1 mixture of **28b** and **29b** resulted in enolization and the formation of a 1:1 mixture of **28b** and **29b** on acidification. The phenyl group makes the α -proton more acidic than those of lactone **12** and hinders the approach of the nucleophile to the carbonyl group. Addition of CeCl₃ to the Grignard reagent helped, but the desired hydroxy vinyl ketone analogous to **13a** was obtained in only 17% yield along with ~70% recovered **28b** and **29b**. Fortunately, treatment of the 6:1 mixture of lactones **28b** and **29b** with *i*-PrMgCl and *N*,*O*-dimethylhydroxylamine-HCl²⁹ afforded Weinreb amides **31a** (78%) and the diastereomer **31b** (13%), which were easily separated. Reaction of **31a** with TESCl, Et₃N, and DMAP gave TES ether **32** in 85% yield, which was treated with vinylmagnesium bromide to give protected vinyl ketone **33** in 72% yield.

The sequence developed to elaborate 13b to 18c-21c worked efficiently to convert 33 to 36 and 37 (see Scheme 8). Cycloaddition of 33 with *N*-SEM-pyrrole oxime and NaOCl afforded a difficultly separable 7:1 mixture of 34 and 35 in 61% yield, whose stereochemistry was assigned from the stereochemistry of the final model spiroiminals 36 and 37. Hydrogenolysis of the isoxazoline over Raney nickel,

Scheme 8. Preparation of Marineosin A Model 36



methylation with NaH and MeI, and HCl-catalyzed hydrolysis of the silyl ether and cyclization afforded the spiroiminals 36 (33% from 34) and 37 (15% from a 1:1 mixture of 34 and 35).

The stereochemistry of 36 and 37 was assigned by analysis of the ¹H NMR spectral data. The CHPh proton H-10 absorbs as a doublet at δ 2.84 (J = 10.9 Hz) in 36 and at δ 3.01 (J = 11.6 Hz) in 37, thereby establishing that both the phenyl and propyl groups are equatorial in both 36 and 37. The axial methyl groups are deshielded by the 1,3-diaxial nitrogen and absorb at δ 1.47 in **36** and δ 1.57 in **37**. These shifts are similar to that of the axial methyl group in marineosin A (1) at δ 1.51 and very different from those of the equatorial methyl groups of 18c, 19c, and 21c at δ 1.13, 1.21, and 1.21, respectively, and the methyl group of marineosin B (2) at δ 1.20. A large NOE between the axial CHPr proton H-9 and the axial methyl group confirmed this stereochemical assignment. The stereochemistry of the methoxy group was established by NOEs between the CHPh proton H-10 and the CHOMe proton H-7 in 36 and between the CHPh methine proton H-10 and the methoxy group in 37 as shown in Scheme 8.

The single stereocenter in 13b is too far from the vinyl ketone to affect the cycloaddition so that 14 was obtained as a 1:1 mixture of isomers. We were very encouraged by the observation that the two additional stereocenters in vinyl ketone 33 are close enough to the double bond to influence the stereochemistry of the cycloaddition leading to 7:1 selectivity favoring the isomer with the marineosin A stereochemistry. Furthermore spiroiminal 36 with a fully substituted tetrahydropyran ring adopts the same conformation as marineosin A, whereas model 20a, which has the same stereochemistry as 36, adopts the other chair conformation with equatorial nitrogen and methyl groups. Minor spiroiminal 37 has the same stereochemistry as marineosin B (2) at the methoxy-substituted carbon, but the opposite stereochemistry at the spiroiminal carbon.

Having developed a practical route to **36** with the marineosin A stereochemistry and conformation, we turned our attention to preparing lactone 7 with the macrocyclic pyrrole tether. *N*-(Ts)-Pyrrole-2-carboxaldehyde (**38**) was treated with 4-pentenylmagnesium bromide, NMO/TPAP, and then NaBH₄ under Muchowski's conditions^{3,30} to generate 2-(5-hexenyl)-pyrrole (**39**)³¹ in 47% overall yield (see Scheme 9). Protection of **39** with (Boc)₂O, Et₃N, and DMAP afforded *N*-Boc-pyrrole

Scheme 9. Synthesis of Macrocyclic Fused Lactone 47b



40 in 74% yield. Boronic acid **41** was prepared using Fürstner's procedure for 4-pentenyl-*N*-Boc-pyrroleboronic acid.³¹ Deprotonation of **40** at the 5-position with lithium 2,2,6,6-tetramethylpiperidide followed by trapping of the resulting carbanion with trimethyl borate gave the unstable boronic acid **41** in about 60% yield. Great care was required during the workup. The organic phases from the extraction were slowly concentrated at room temperature until a solid started to precipitate. The mixture was then cooled to 0 °C and was filtered. Trituration of the solid with cold ether afforded **41** as a yellowish solid that was used immediately for the Suzuki coupling. To our delight, treatment of iodolactone **11** and boronic acid **41** with Pd(PPh₃)₄, Na₂CO₃, and LiCl in aqueous 1,2-dimethoxyethane at 80 °C afforded pyrrolyl lactone **42** in 55% yield.

We were unable to remove the Boc protecting group at this point, so we treated unsaturated lactone **42** with allylmagnesium bromide, ZnBr₂, and TMSCI. To our surprise, we obtained a difficult to separate 5:4 mixture of the undesired *cis* isomer **43** and ketene acylal **44** resulting from Boc migration in 56% yield. The CH-pyrrole proton of **43** absorbs as a doublet at δ 4.73 (J = 4.9 Hz) analogously to that of **29a**, δ 3.94 (J = 5.6Hz). There is no CH-pyrrole proton in **44**, but LC-MS analysis indicates that it has the same molecular weight as **43**, suggesting that a precedented Boc migration occurred.³² Apparently, the enolate generated by conjugate addition of the allyl group undergoes kinetically controlled protonation from the face opposite the allyl group to give *cis* isomer **43** and Boc migration to give **44**. Fortunately, treatment of the 5:4 mixture of **43** and **44** with TMSOTf and 2,6-lutidine^{32a} removed the Boc groups from both compounds and epimerized the α position to afford a 19:1 mixture of the desired deprotected *trans* isomer 45 and the deprotected *cis* isomer in 91% yield. The CH-pyrrole proton of 45 absorbs as a doublet at δ 3.57 (J = 6.7 Hz) and shows a strong NOE to the CHMe proton, establishing that lactone 45 also adopts a boat conformation. The coupling constant is smaller than those of the CHPh proton (J = 9.8 Hz) in 28a and the CH-N-SEMpyrrole proton (J = 9.2 Hz) in the protected analogue of 45 that leads to 47a. This suggests that hydrogen bonding between the lactone carbonyl group and the pyrrole nitrogen perturbs the lactone conformation in 45.

Treatment of a 10^{-4} molar solution of 45 with 15 mol % Grubbs II catalyst in CH₂Cl₂ at reflux for 12 h gave 46 as a *cis/* trans mixture of isomers in 18% yield. The yield of 46 was increased to 41% by using more catalyst $(2 \times 15 \text{ mol } \%)$ and prolonging the reaction time to 16 h. Although this yield was not satisfactory, it provided sufficient material for further elaboration. Hydrogenation of 46 over Raney nickel reduced the double bond to afford 47b as a single compound in 90% yield. The ¹H NMR spectrum of 47b shows the expected shielding of protons on the seven-carbon methylene bridge by the pyrrole ring to δ 0.83 (m, 1) and δ 0.39 (m, 1), similar to that observed in marineosins A at δ 0.69 (m, 1) and δ 0.52 (m, 1). The large coupling constant for the CH-pyrrole proton at δ 3.47 (J = 12.1 Hz) indicates that the macrocycle is *trans* fused to the lactone. The structure of 47b was confirmed by X-ray crystal structure determination, which indicates that the lactone adopts the boat conformation as shown in Figure 1.



Figure 1. Structure of **47b** determined by X-ray crystallography. The tether is disordered with two positions for atom C10; only the major component (92.2%) is shown.

N-H-Pyrrole lactone **47b** was converted to *N*-H-pyrrole isoxazoline **51b** by the sequence developed to prepare isoxazolines **34** and **35** (see Scheme 10). Treatment of **47b** with *i*-PrMgCl and *N*,*O*-dimethylhydroxylamine-HCl afforded Weinreb amide **48b** in 86% yield, which was protected to give triethylsilyl ether **49b** in 77% yield. To our surprise, considerable reduction to the aldehyde occurred on treatment of Weinreb amide **49b** with vinylmagnesium bromide. This problem has been previously noted, especially as the Grignard reagent ages.^{33,34} Fortunately, addition of a solution of vinyllithium³⁴ freshly prepared from vinyl bromide and *n*-BuLi to **49b** afforded enone **50b** in 70% yield.

The cycloaddition of **50b** with N-SEM-pyrrole-2-carboxaldehyde oxime and NaOCl gave isoxazoline **51b** in \sim 67% yield as





a mixture of stereoisomers, although the reaction was not as clean as those with enones 13b and 33. Hydrogenolysis of 51b over Raney nickel provided a crude mixture in ~67% yield that appeared to contain hemi-iminals 52b based on the similarity of the ¹H NMR spectrum to those of 15 and the crude hydrogenation product from 34.

We were concerned that it might not be possible to methylate the two hydroxy groups of **52b** without *N*methylation of the pyrrole. However, to our disappointment, *O*-methylation of **52b**, with or without concomitant *N*methylation, could not be accomplished. The use of NaH and MeI, which was successful in all of the model studies, afforded a complex mixture. Other bases (KOH or KOtBu) and methylating reagents (Me₂SO₄) were also successful with **15a** as was acid-catalyzed methylation with CH_2N_2 or TMSCHN₂. Unfortunately, all of these conditions failed to methylate hemiiminal **52b**. Hemi-iminals are unstable and may decompose if the methylation does not occur rapidly. Apparently the hydroxy groups of **52b** are more hindered than those of the model hemi-iminals so that only decomposition occurs on attempted methylation.

We attempted to form the spiroiminal prior to methylation, but **52b** decomposed on treatment with 2 M HCl or Dowex 50 WX ion-exchange resin. Hemi-iminal **52b** decomposed after 10 h in CDCl₃, indicating that it is very acid-sensitive.

We had previously prepared macrocyclic *N*-SEM pyrrole lactone 47a.^{13,35} The four-step sequence leading to isoxazoline **51a** proceeded uneventfully, but all attempts to hydrogenolyze the isoxazoline and to form hemi-iminal **52a** failed completely. We hypothesized that the protecting group on the pyrrole in the tether prevented formation of the hemi-iminal. We could not deprotect 47a to give 47b, so we developed the route to **47b** described in detail above. We can form hemi-iminal **52b** lacking the pyrrole protecting group, but the sequence fails one step later at the methylation stage.

In conclusion, we have developed a short and efficient synthesis of model spiroiminals 18a-21a (six steps) and 18b-21b (seven steps) that have the same stereochemistry as marineosins A and B, but different conformations. Phenyl-substituted spiroiminals 18a-21a were also prepared biomimetically by reduction of an enol ether. More highly substituted spiroiminal 36 with the same stereochemistry and

conformation as marineosin A was prepared in 11 steps. Macrocyclic pyrrole lactone **47b** was prepared stereospecifically in 10 steps. A five-step sequence converted the lactone to a late hemi-iminal intermediate **52b** that has resisted the methylation and spiroiminal formation that would lead to marineosin A.

EXPERIMENTAL SECTION

General Experimental Methods. Reactions were conducted in flame- or oven-dried glassware under a nitrogen atmosphere and were stirred magnetically. The phrase "concentrated" refers to removal of solvents by means of a rotary evaporator attached to a diaphragm pump (15-60 Torr) followed by removal of residual solvents at <1 Torr with a vacuum pump. Flash chromatography was performed on silica gel 60 (230-400 mesh). Analytical thin layer chromatography (TLC) was performed using silica gel 60 F-254 precoated glass plates (0.25 mm). TLC plates were analyzed by short wave UV illumination or by dipping in vanillin stain (27 g of vanillin in 380 mL of EtOH, 50 mL of water, and 20 mL of concentrated sulfuric acid) and heating on a hot plate. THF and ether were dried and purified by distillation from sodium/benzophenone. Et₃N was distilled from CaH₂. ¹H and ¹³C NMR spectra were obtained on a 400 MHz spectrometer in CDCl₃ with CHCl₃ as an internal standard (δ 7.26, CDCl₃ at δ 77.00) unless otherwise indicated. Chemical shifts are reported in δ (ppm downfield from tetramethylsilane). Coupling constants are reported in hertz with multiplicities denoted as s (singlet), d (doublet), t (triplet), q (quartet), p (pentet), m (multiplet), and br (broad). IR spectra were acquired on an FT-IR spectrometer and are reported in wave numbers (cm^{-1}) . High resolution mass spectra were obtained using the following ionization techniques: chemical ionization (CI), electron impact (EI), electrospray ionization analyzed by quadrupole time-offlight (QTOF).

Benzaldehyde Oxime. A solution of benzaldehyde (530 mg, 5.0 mmol) in 20 mL of EtOH was treated with a mixture of NaOH (300 mg, 7.50 mmol) and NH₂OH·HCl (783 mg, 11.4 mmol) in 10 mL of H₂O. The reaction mixture was stirred at 25 °C for 6 h, concentrated to remove EtOH, diluted with CH_2Cl_2 , washed with brine, and dried (Na₂SO₄). Flash chromatography on silica gel (8:1 hexanes/EtOAc) gave 482 mg (84%) of the oxime as a brown solid with data identical to those previously reported.⁴¹

1-[[2-(Trimethylsilyl)ethoxy]methyl]-1H-pyrrole-2-carboxaldehyde. Protection of the pyrrole was carried out by the literature procedure.¹⁸ A solution of pyrrole-2-carboxaldehyde (245 mg, 2.57 mmol) in anhydrous THF (2 mL) was added dropwise to a suspension of NaH (60% in mineral oil, 124 mg, 3.09 mmol) in THF (10 mL) at 0 °C. The mixture was stirred at 0 °C for 30 min, and SEMCl (0.50 mL, 2.83 mmol) was added by syringe over 3 min. The reaction was warmed to 25 °C and stirred for 2 h. The mixture was quenched with saturated aqueous NH₄Cl (3 mL). The aqueous layer was extracted with EtOAc, and the combined organic layers were dried (Na₂SO₄) and concentrated. Flash chromatography on silica gel (8:1 hexanes/EtOAc) gave 538 mg (93%) of N-SEM-pyrrole-2-carboxaldehyde as a pale yellow gum: ¹H NMR 9.58 (s, 1), 7.15-7.13 (m, 1), 6.96 (dd, 1, J = 1.5, 3), 6.30 (dd, 1, J = 3, 4), 5.70 (s, 2), 3.54 (t, 2, J = 8.1), 0.89 (t, 2, J = 8.1), -0.04 (s, 9); ¹³C NMR 179.3, 131.6, 130.8, 125.0, 110.2, 76.2, 65.8, 17.5, -1.7 (3 C); IR (neat) 1671.

1-[[2-(Trimethylsilyl)ethoxy]methyl]-1*H***-pyrrole-2-carboxaldehyde Oxime.** A solution of *N*-SEM-pyrrole-2-carboxaldehyde (538 mg, 2.39 mmol) in 11 mL of 10:1 MeOH/H₂O was treated with NH₂OH-HCl (183 mg, 2.63 mmol) and NaOAc (295 mg, 3.59 mmol). The resulting mixture was stirred at 25 °C for 2.5 h, concentrated to remove MeOH, diluted with CH₂Cl₂, washed with brine, dried (Na₂SO₄), and concentrated. Flash chromatography on silica gel (8:1 hexanes/EtOAc) gave 482 mg (84%) of the oxime as a brown gum: ¹H NMR 8.92 (s, 1, OH), 8.20 (s, 1), 6.87–6.85 (m, 1), 6.52 (dd, 1, *J* = 1.2, 2.5), 6.19 (dd, 1, *J* = 3, 4), 5.46 (s, 2), 3.50 (t, 2, *J* = 8.2), 0.91 (t, 2, *J* = 8.2), -0.03 (s, 9); ¹³C NMR 142.3, 126.2, 125.2, 115.0, 109.1, 76.9, 65.5, 17.5, -1.6 (3 C); IR (neat) 3376, 1624; HRMS (EI) calcd for C₁₁H₂₀N₂O₂Si (M⁺) 240.1294, found 240.1298. **7-Hydroxy-1-octen-3-one (13a).** Enone **13a** was prepared by the literature procedure.¹¹ A solution of 6-methyltetrahydropyran-2-one (**12**) (0.92 g, 8.76 mmol) in anhydrous THF (15 mL) was treated with vinylmagnesium bromide (1 M in THF, 10.51 mL, 10.51 mmol) by syringe over 15 min under nitrogen at -78 °C. The resulting solution was stirred at -78 °C for 4 h. The mixture was quenched with saturated aqueous NH₄Cl, diluted with EtOAc, washed with brine, dried (Na₂SO₄), and concentrated to give 1.11 g of crude **13a**. Flash chromatography on MeOH-deactivated silica gel (4:1 hexanes/EtOAc) gave 1.06 g (85%) of **13a** as a colorless liquid: ¹H NMR 6.36 (dd, 1, *J* = 10.4, 17.4), 6.24 (d, 1, *J* = 17.4), 5.85 (d, 1, *J* = 10.4), 3.81–3.76 (m, 1), 2.64 (t, 2, *J* = 6.7), 2.37 (s, 1, OH), 1.75–1.65 (m, 2), 1.50–1.43 (m, 2), 1.19 (d, 3, *J* = 6.7); ¹³C NMR 201.0, 136.3, 128.2, 67.3, 39.2, 38.4, 23.3, 19.8; IR (neat) 3452 (br), 1729.

7-Triethylsilyloxy-1-octen-3-one (13b). A solution of alcohol **13a** (836 mg, 5.88 mmol) in 15 mL of THF was treated with Et₃N (1.36 mL, 9.41 mmol), DMAP (69 mg, 0.59 mmol), and TESCl (1.58 mL, 9.41 mmol). The mixture was stirred at 25 °C for 3 h. The reaction was then diluted with Et₂O (10 mL) and washed with brine (3 × 5 mL). The organic layer was dried (MgSO₄) and concentrated to give 1.78 g of crude **13b**. Flash chromatography on silica gel (18:1 hexanes/EtOAc) gave 1.45 g (96%) of **13b** as a sticky liquid: ¹H NMR 6.34 (dd, 1, *J* = 10.6, 17.6), 6.21 (d, 1, *J* = 17.6), 5.81 (d, 1, *J* = 10.6), 3.82–3.78 (m, 1), 2.59 (t, 2, *J* = 6.4), 1.72–1.58 (m, 2), 1.46–1.39 (m, 2), 1.14 (d, 3, *J* = 6.4), 0.95 (t, 9, *J* = 7.6), 0.58 (q, 6, *J* = 7.6); ¹³C NMR 200.8, 136.5, 127.9, 68.2, 39.6, 39.1, 23.8, 20.3, 6.9 (3 C), 4.9 (3 C); IR (neat) 1682; HRMS (EI) calcd for C₁₄H₂₇O₂Si (M – H⁺) 255.1780, found 255.1787.

1-(4,5-Dihydro-3-phenyl-5-isoxazolyl)-5-triethylsilyloxy-1hexanone (14a). A solution of N-chlorosuccinimide (220 mg, 1.65 mmol) in anhydrous THF (3 mL) was added dropwise by syringe over 20 min to a solution of benzaldehyde oxime (170 mg, 1.40 mmol) in THF (6 mL). The mixture was stirred at 25 °C for 5 h, cooled to -78°C, and treated with a solution of enone 13b (300 mg, 1.17 mmol) in THF (2 mL) and then Et₃N (240 μ L, 1.65 mmol). The mixture was gradually warmed to 25 °C and stirred for 3 h. The reaction mixture was diluted with EtOAc, washed with brine, dried (Na2SO4), and concentrated. Flash chromatography on MeOH-deactivated silica gel (12:1 hexanes/EtOAc) gave 341 mg (78%) of 14a as a 1:1 mixture of diastereomers as a colorless gum: ¹H NMR 7.67 (d, 2, J = 6.1), 7.43– 7.39 (m, 3), 5.03 (dd, 1, J = 6.1, 12.1), 3.79 (tq, 1, J = 6.1, 6.1), 3.64 (dd, 1, J = 6.1, 16.8), 3.48 (dd, 1, J = 12.1, 16.8), 2.73 (t, 2, J = 7.3),1.73–1.52 (m, 2), 1.50–1.34 (m, 2), 1.12 (d, 3, J = 5.5), 0.94 (t, 9, J = 6.6), 0.57 (q, 6, J = 6.6); ¹³C NMR 209.5, 156.6, 130.5, 128.8 (2 C), 128.5, 126.8 (2 C), 84.1, 68.1, (38.96, 38.94), (38.82, 38.81), (37.28, 37.25), 23.7, (19.30, 19.27), 6.8 (3 C), 4.9 (3 C); IR (neat) 1721, 1595; HRMS (EI) calcd for C₁₉H₂₈O₃NSi (M⁺ - CH₂CH₃) 346.1838, found 346.1837.

1-[4,5-Dihydro-3-[1-[[2-(trimethylsilyl)ethoxy]methyl]-1Hpyrrol-2-yl]-5-isoxazolyl]-5-triethylsilyloxy-1-hexanone (14c). A mixture of N-SEM-pyrrole-2-carboxaldehyde oxime (440 mg, 1.83 mmol) and enone 13b (610 mg, 2.28 mmol) in CH₂Cl₂ (15 mL) was treated with bleach (5.25% aqueous NaOCl, 5.15 mL, 271 mg of NaOCl, 3.66 mmol) and Et₃N (40 μ L, 0.28 mmol) at 0 °C. The resulting mixture was warmed to 25 $\,^{\circ}\text{C}$ and stirred for 3 h. The reaction was then diluted with CH2Cl2, washed with brine, dried (Na₂SO₄), and concentrated. Flash chromatography on MeOHdeactivated silica gel (12:1 hexanes/EtOAc) gave 661 mg (73%) of 14c as a mixture of diastereomers as a pale yellow gum: ¹H NMR 7.00–6.98 (m, 1), 6.46–6.44 (m, 1), 6.23–6.12 (m, 1), 5.67 (d, 1, J = 10.4), 5.60 (d, 1, J = 10.4), 4.87 (dd, 1, J = 6.2, 11.3), 3.79 (tq, 1, J = 6.1, 6.1, 3.60 (dd, 1, I = 6.2, 16.3), 3.53 - 3.46 (m, 3), 2.78 - 2.63 (m, 2), 1.72–1.50 (m, 2), 1.48–1.34 (m, 2), 1.12 (d, 3, *J* = 6.1), 0.94 (t, 9, J = 7.8, 0.89 (t, 2, J = 7.9), 0.57 (q, 6, J = 7.8), -0.04 (s, 9); ¹³C NMR 209.7, 150.1, 127.5, 121.3, 116.0, 109.3, 82.4, 77.4, 68.1, 65.7, 39.4, 39.0, 38.8, (23.72, 23.70), 19.3, 17.7, 6.9 (3 C), 4.9 (3 C), -1.5 (3 C); IR (neat) 1721, 1598; HRMS (EI) calcd for $C_{25}H_{46}O_4N_2Si_2~(M^{\scriptscriptstyle +})$ 494.2996, found 494.2989.

3,4-Dihydro-2,3-dimethoxy-2-(4-triethylsilyloxypentyl)-4phenyl-2H-pyrrole (16a). A solution of isoxazoline 14a (178 mg, 0.47 mmol) in 10 mL of MeOH was treated with a wet slurry of Raney nickel 2800 (~50 mg), and the suspension was stirred at 25 °C under H₂ (1 atm) for 35 min. The mixture was then diluted with EtOAc and filtered. The filtrate was washed with brine (3 \times 5 mL), dried (MgSO₄), and concentrated to give 174 mg of crude hydroxy hemiiminal **15a** as a mixture of four diastereomers that was used for the next step.

A solution of crude 15a in anhydrous THF (2 mL) was added dropwise to a suspension of NaH (60% in mineral oil, 152 mg, 3.80 mmol) in THF (5 mL) at 0 °C. The mixture was stirred at 0 °C for 30 min, and MeI (237 μ L, 3.80 mmol) was then added by syringe over 3 min. The resulting mixture was warmed to 25 °C and stirred for 4 h. The reaction was quenched with saturated aqueous NH₄Cl (3 mL). The aqueous layer was extracted with EtOAc (3×5 mL). The combined organic layers were dried (Na2SO4) and concentrated to give 163 mg of crude 16a. Flash chromatography on silica gel (18:1 hexanes/EtOAc) gave 110 mg (58% for two steps) of 16a as a mixture of four diastereomers as a colorless gum: ¹H NMR (major (75–80%) pair of diastereomers with either cis or trans methoxy groups) 7.87 (d, 2, J = 7.3), 7.46-7.39 (m, 3), 3.96-3.91 (m, 1), 3.84-3.75 (m, 1), 3.48 (s, 6), 3.20 (dd, 1, J = 7.3, 17.4), 3.02 (dd, 1, J = 3.0, 17.4), 1.96-1.82 (m, 1), 1.67–1.35 (m, 5), 1.14 (d, 3, *J* = 6.1), 0.93 (t, 9, *J* = 7.8), 0.58 (q, 6, J = 7.8); ¹H NMR (minor (20-25%) pair of diastereomers with either trans or cis methoxy groups) 3.48-3.24 (m, 2 or 3); IR (neat) 2955, 1619, 1449; HRMS (EI) calcd for C₂₃H₃₉O₃NSi (M⁺) 405.2699. found 405.2710.

3,4-Dihydro-2,3-dimethoxy-2-(4-triethylsilyloxypentyl)-4-(1-[[2-(trimethylsilyl)ethoxy]methyl]-1*H*-pyrrol-2-yl)-2*H*-pyrrole (16c). A solution of isoxazoline 14c (203 mg, 0.41 mmol) in 12 mL of 5:1 MeOH/H₂O was treated with a wet slurry of Raney nickel 2800 (~50 mg), and the suspension was stirred at 25 °C under H₂ (1 atm) for about 50 min. The mixture was then diluted with EtOAc and filtered. The filtrate was washed with brine (3 × 5 mL), dried (Na₂SO₄), and concentrated to give 191 mg of crude hydroxy hemiiminal 15c.

A solution of crude 15c in THF (2 mL) was added dropwise to a suspension of NaH (60% in mineral oil, 130 mg, 3.24 mmol) in THF (5 mL) at 0 °C. The mixture was stirred at 0 °C for 30 min, and MeI (203 μ L, 3.24 mmol) was added dropwise by syringe over 3 min. The resulting mixture was warmed to 25 °C and stirred for 4 h. The reaction was quenched with saturated aqueous NH₄Cl (3 mL), and the aqueous layer was extracted with EtOAc. The combined organic layers were dried (Na₂SO₄) and concentrated to give 151 mg of crude 16c. Flash chromatography on silica gel (15:1 hexanes/EtOAc) gave 91 mg (42% for two steps) of 16c (pale yellow gum) as a mixture of four diastereomers in which two predominate: ¹H NMR 7.03-7.01 (m, 1), 6.58-6.56 (m, 1), 6.21-6.19 (m, 1), 5.93 (d, 1, J = 10.4), 5.90 (d, 1, J = 10.4), 3.80-3.77 (m, 2), 3.54 (t, 2, J = 7.9), 3.44 (s, 3), 3.43 (s, 3), 3.12 (dd, 1, *J* = 6.7, 17.1), 2.96 (dd, 1, *J* = 2.4, 17.1), 1.83–1.77 (m, 1), 1.55–1.37 (m, 5), 1.13 (d, 3, *J* = 6.1), 0.94 (t, 9, *J* = 7.8), 0.87 (t, 2, *J* = 7.9), 0.57 (q, 6, J = 7.8), -0.05 (s, 9); IR (neat) 2954, 1617; HRMS (EI) calcd for C₂₇H₅₂O₄N₂Si₂ (M⁺) 524.3466, found 524.3475.

3-Methoxy-α-methyl-5-phenyl-1*H*-pyrrole-2-butanol (17a) and (±)-(4*S*,5*R*,7*R*)-, (±)-(4*R*,5*R*,7*R*)-, and (±)-(4*R*,5*S*,7*R*)-4-**Methoxy-7-methyl-2-phenyl-6-oxa-1-azaspiro**[4.5]dec-1-ene (18a, 19a, and 21a). A solution of 16a (101 mg, 243 µmol) in 6 mL of 1:1 CH₃CN/THF was treated with 2 M HCl (2.49 mL, 4.98 mmol) at 0 °C. The resulting mixture was stirred at 0 °C for 40 min. Saturated NaHCO₃ (5 mL) was added to bring the pH to 7. The reaction was extracted with EtOAc, and the organic layer was washed with brine, dried (Na₂SO₄), and concentrated. Flash chromatography on MeOHdeactivated silica gel (7:1 to 2:1 hexanes/EtOAc) gave 26 mg (41%) of 18a as a colorless gum, followed by 8.1 mg (13%) of 19a as a colorless gum, 7.4 mg (12%) of 21a as a colorless gum, and then 5.2 mg (8%) of 17a as a pale yellow gum.

Data for 17a: ¹H NMR (recorded in C_6D_6 because the compound is unstable in $CDCl_3^{21}$) 7.83 (br, 1, NH), 7.30 (d, 2, *J* = 7.3), 7.20 (t, 2, *J* = 7.3), 7.04 (t, 1, *J* = 7.3), 6.30 (d, 1, *J* = 2.5), 3.60 (s, 3), 3.58–3.48 (m, 1), 2.61 (t, 2, *J* = 7.4), 1.69–1.51 (m, 2), 1.38–1.25 (m, 2), 0.92 (d, 3, *J* = 6.1); ¹³C NMR (C_6D_6) 146.0, 133.9, 129.0 (2 C), 126.9,

125.5, 123.4 (2 C), 117.5, 95.0, 67.8, 58.5, 38.3, 26.3, 24.5, 24.0; IR (neat) 3316, 2934, 1630; HRMS (EI) calcd for $C_{16}H_{19}O_2N$ (M⁺-2H) 257.1416, found 257.1407. This compound is unstable and oxidizes easily to **24** and **25**.

Data for 18a: ¹H NMR 7.84 (d, 2, J = 6.7), 7.44–7.38 (m, 3), 4.43– 4.39 (m, 1, H-7), 3.88 (dd, 1, J = 7.0, 7.0, H-4), 3.46 (s, 3), 3.30 (dd, 1, J = 17.1, 7.0, H-3), 2.77 (dd, 1, J = 16.4, 7.0, H-3), 2.07 (br ddd, 1, J =11, 11, 11, H-9ax), 1.79 (ddd, 1, J = 11, 11, 3, H-10ax), 1.77–1.69 (m, 2, H-8eq, H-9eq), 1.51 (br d, 1, J = 11, H-10eq), 1.36 (br ddd, 1, J =11, 11, 11, H-8ax), 1.16 (d, 3, J = 6.1, H-7 Me); ¹³C NMR 169.6, 134.8, 130.8, 128.4 (2 C), 127.6 (2 C), 103.8, 87.2, 68.7, 58.2, 39.1, 33.6, 28.7, 22.4, 19.8; IR (neat) 2932, 1615, 1448; HRMS (EI) calcd for C₁₆H₂₁O₂N (M⁺) 259.1572, found 259.1560. A 1D NOESY experiment with irradiation of H-4 at δ 3.88 showed NOEs to OMe at δ 3.46 (OMe) and H-3s at δ 3.30 and 2.77. A 1D NOESY experiment with irradiation of H-7 at δ 4.43–4.39 showed NOEs to H-9ax at δ 2.07, H-8eq at δ 1.77–1.69, and 7-Me at δ 1.16.

Data for **19a**: ¹H NMR 7.86 (d, 2, *J* = 7.3), 7.46–7.36 (m, 3), 4.49– 4.42 (m, 1, H-7), 3.77 (dd, 1, *J* = 6.1, 4.0, H-4), 3.50 (s, 3), 3.13 (dd, 1, *J* = 17.1, 6.1, H-3), 3.05 (dd, 1, *J* = 17.1, 4.0, H-3), 2.12 (br ddd, 1, *J* = 11, 11, 11, H-9ax), 1.77 (ddd, 1, *J* = 11, 11, 3, H-10ax), 1.76–1.68 (m, 2, H-8eq, H-9eq), 1.48 (br d, 1, *J* = 11, H-10eq), 1.40 (br ddd, 1, *J* = 11, 11, 11, H-8ax), 1.23 (d, 3, *J* = 6.7, H-7 Me); ¹³C NMR 170.3, 134.8, 130.7, 128.3 (2 C), 127.7 (2 C), 101.7, 85.4, 68.6, 58.8, 39.6, 34.7, 33.3, 22.3, 20.4; IR (neat) 2930, 1616, 1448; HRMS (EI) calcd for C₁₆H₂₁O₂N (M⁺) 259.1572, found 259.1570. A 1D NOESY experiment with irradiation of H-4 at δ 3.77 showed NOEs to OMe at δ 3.50 (OMe), H-3s at δ 3.13 and 3.05, and H-10ax at δ 1.77 and H-10eq at δ 1.48.

Data for **21a**: ¹H NMR 7.90 (d, 2, J = 7.4), 7.44–7.36 (m, 3), 4.11 (dd, 1, J = 6.7, 3.6, H-4), 3.83–3.76 (m, 1, H-7), 3.40 (s, 3), 3.36 (dd, 1, J = 17.4, 6.7, H-3), 2.94 (dd, 1, J = 17.4, 3.6, H-3), 2.06–2.01 (m, 1), 1.88–1.75 (m, 3), 1.62 (br d, 1, J = 11), 1.46–1.38 (m, 1), 1.23 (d, 3, J = 6.1); ¹³C NMR 171.9, 134.0, 131.1, 128.2 (2 C), 128.1 (2 C), 105.4, 83.8, 69.9, 57.7, 40.1, 32.2, 28.9, 22.3, 20.6; IR (neat) 2930, 1627, 1448; HRMS (EI) calcd for C₁₆H₂₁O₂N (M⁺) 259.1572, found 259.1556. A 1D NOESY experiment with irradiation of H-4 at δ 4.11 showed NOEs to H-7 at δ 3.83–3.76, OMe at δ 3.40, and H-3s at δ 3.36 and 2.94.

Equilibration of 19a and 21a. A solution of **19a** in 0.6 mL of $CDCl_3$ (containing HCl/DCl from decomposition of $CDCl_3$) equilibrated to a 3:1 mixture of **19a** and **21a**. The percentage of **19a** in the mixture was determined as a function of time by ¹H NMR spectroscopy: initial, 100%; 7 days, 90%; 14 days, 80%; 20 days, 75%. The spectrum did not change at longer times. A solution of **21a** in 0.6 mL of $CDCl_3$ (containing HCl/DCl from decomposition of $CDCl_3$) equilibrated to a 3:1 mixture of **19a** and **21a**. The percentage of **19a** in the mixture was determined as a function of time by ¹H NMR spectroscopy: initial, <20; 5 days, 25%; 10 days, 60%, 15 days, 75%. The spectrum did not change at longer times.

Equilibration of 18a and 20a. A solution of **18a** in 0.6 mL of CDCl₃ (containing HCl/DCl from decomposition of CDCl₃) was monitored by ¹H NMR for 14 days, at which time a 19:1 mixture of **18a** and **20a** was present. Partial data for **20a** were determined from the mixture: ¹H NMR 7.91 (d, 2, J = 7.6), 7.40–7.20 (m, 3), 4.13 (d, 1, J = 4.9, H-4), 3.80–3.74 (m, 1, H-7), 3.36 (s, 3, OMe), 3.20 (d, 1, J = 17.4, H-3), 2.99 (dd, 1, J = 17.4, 4.9, H-3), 1.30 (d, 3, J = 6.4).

(±)-(4*X*,5*R*,7*R*)-, (±)-(4*R*,5*R*,7*R*)-, and (±)-(4*R*,5*S*,7*R*)-4-Methoxy-7-methyl-2-(1-[[2-(trimethylsilyl)ethoxy]methyl]-1*H*-pyrrol-2-yl)-6-oxa-1-azaspiro[4.5]dec-1-ene (18c, 19c, and 21c). A solution of 16c (78 mg, 149 µmol) in 8 mL of 3:1 CH₃CN/THF was treated with aqueous 2 M HCl (1.49 mL, 2.98 µmol) at 25 °C. The resulting mixture was stirred at 25 °C for 11 h. Saturated NaHCO₃ (3 mL) was added to bring the pH to 7. The reaction was extracted with EtOAc, and the organic layer was washed with brine, dried (Na₂SO₄), and concentrated to give 77 mg of a mixture of spiroiminals. Flash chromatography on MeOH-deactivated silica gel (18:1 to 2:1 hexanes/ EtOAc) gave 19 mg (34%) of isomer 18c as a colorless gum followed by 19 mg (34%) of an inseparable 3:2 mixture of isomers 19c and 21c as a colorless gum. Data for **18c**: ¹H NMR 7.02–7.00 (m, 1), 6.57–6.55 (m, 1), 6.21– 6.19 (m, 1), 6.01 (d, 1, J = 10.1), 5.88 (d, 1, J = 10.1), 4.29–4.22 (m, 1, H-7), 3.77 (dd, 1, J = 6.9, 6.9, H-4), 3.55 (t, 2, J = 8.2), 3.43 (s, 3), 3.23 (dd, 1, J = 6.9, 16.3, H-3), 2.76 (dd, 1, J = 6.9, 16.3, H-3), 1.98 (br ddd, 1, J = 11, 11, 11, H-9ax), 1.76 (ddd, 1, J = 11, 11, 3, H-10ax), 1.76– 1.64 (m, 2, H-8eq, H-9eq), 1.49 (br d, 1, J = 11, H-10eq), 1.34 (br ddd, 1, J = 11, 11, 11, H-8ax), 1.13 (d, 3, J = 6.1), 0.88 (t, 2, J = 8.2), -0.05 (s, 9); ¹³C NMR 162.6, 127.6, 127.5, 116.6, 108.9, 104.3, 86.2, 76.8, 68.6, 65.5, 58.2, 40.5, 33.6, 29.0, 22.4, 20.0, 18.0, -1.5 (3 C); IR (neat) 1610; HRMS (EI) calcd for C₂₀H₃₄O₃N₂Si (M⁺) 378.2339, found 378.2325.

Data for **19c** and **21c**: IR (CDCl₃) 1613; HRMS (EI) calcd for $C_{20}H_{34}O_3N_2Si$ (M⁺) 378.2339, found 378.2350.

NMR data for **19c** were determined from the mixture: ¹H NMR 7.03–7.01 (m, 1), 6.57–6.55 (m, 1), 6.20–6.17 (m, 1), 6.16 (d, 1, *J* = 10.1), 5.74 (d, 1, *J* = 10.1), 4.35–4.29 (m, 1), 3.67 (dd, 1, *J* = 6.0, 4.8), 3.56 (t, 2, *J* = 8.5), 3.47 (s, 3), 3.06 (dd, 1, *J* = 17.2, 6.0), 3.01 (dd, 1, *J* = 17.2, 4.8), 2.06–1.99 (m, 1), 1.78–1.35 (m, 5), 1.21 (d, 3, *J* = 6.1), 0.90–0.85 (m, 2), -0.05 (s, 9); ¹³C NMR 162.9, 127.6, 127.4, 116.7, 108.9, 102.0, 84.3, 68.6, 65.6, 58.6, 40.8, 34.8, 33.4, 22.3, 20.6, 18.0, -1.5 (3 C), (one peak is obscured by the CDCl₃ triplet at δ 77.0).

NMR data for **21c** were determined from the mixture: ¹H NMR 7.01–6.99 (m, 1), 6.57–6.55 (m, 1), 6.38 (d, 1, J = 10.4), 6.20–6.17 (m, 1), 5.54 (d, 1, J = 10.4), 3.97 (dd, 1, J = 6.0, 3.1), 3.81–3.75 (m, 1), 3.51 (t, 2, J = 8.5), 3.36 (s, 3), 3.28 (dd, 1, J = 16.8, 6.0), 2.86 (dd, 1, J = 16.8, 3.1), 2.06–1.99 (m, 1), 1.78–1.35 (m, 5), 1.21 (d, 3, J = 6.1), 0.90–0.85 (m, 2), -0.04 (s, 9) ¹³C NMR 164.8, 127.6, 127.4, 117.0, 108.9, 106.2, 82.0, 70.1, 65.6, 57.4, 41.2, 32.2, 29.4, 22.4, 20.5, 17.9, -1.5 (3 C), (one peak is obscured by the CDCl₃ triplet at δ 77.0).

(+)-(4S,5R,7R)-4-Methoxy-7-methyl-2-(1H-pyrrol-2-yl)-6oxa-1-azaspiro[4.5]dec-1-ene (18b). A mixture of 18c (19 mg, 50.2 μ mol) and molecular sieves (4 Å, 100 mg) in freshly distilled THF (3 mL) was treated with TBAF (1 M in THF, 1.01 mL, 1.01 mmol) dropwise at 50 °C. The resulting mixture was stirred at 60 °C for 3 h. The reaction was cooled, diluted with Et₂O (15 mL), and washed with brine $(2 \times 5 \text{ mL})$ and H₂O $(3 \times 5 \text{ mL})$. The organic layer was dried (MgSO₄) and concentrated to give 59 mg of crude 18b. Flash chromatography on MeOH-deactivated silica gel (4:1 hexanes/ EtOAc) gave 5.7 mg (54%) of 18b as a pale yellow gum: ¹H NMR 6.94-6.91 (m, 1), 6.57-6.54 (m, 1), 6.25-6.23 (m, 1), 4.26-4.20 (m, 1), 3.82 (dd, 1, *J* = 6.7, 6.1), 3.43 (s, 3), 3.19 (dd, 1, *J* = 16.4, 6.7), 2.73 (dd, 1, *J* = 16.4, 6.1), 1.97 (br ddd, 1, *J* = 11, 11, 11, H-9ax), 1.81–1.66 (m, 3, H-10ax, H-8eq, H-9eq), 1.54 (br d, 1, J = 11, H-10eq), 1.32 (br ddd, 1, *J* = 11, 11, 11, H-8ax), 1.12 (d, 3, *J* = 6.1, H-7 Me), the pyrrole NH was not observed; ¹³C NMR 162.8, 127.7, 122.1, 113.7, 109.8, 103.4, 86.9, 68.5, 58.1, 38.5, 33.4, 28.7, 22.4, 19.7; IR (CDCl₃) 2930, 1607, 1432, 743; HRMS (EI) calcd C₁₄H₂₀N₂O₂ (M⁺) 248.1525, found 248.1532.

(±)-(4*R*,5*R*,7*R*)- and (±)-(4*R*,55,7*R*)-4-Methoxy-7-methyl-2-(1*H*-pyrrol-2-yl)-6-oxa-1-azaspiro[4.5]dec-1-ene (19b and 21b). A 3:2 mixture of 19c and 21c (19 mg, 50.2 μ mol), and molecular sieves (4 Å, 100 mg) in freshly distilled THF (3 mL) was treated with TBAF (1 M in THF, 1.01 mL, 1.01 mmol) dropwise at 50 °C. The resulting mixture was then stirred at 60 °C for 3 h. The reaction was cooled, diluted with Et₂O (15 mL), and washed with brine (2 × 5 mL) and H₂O (3 × 5 mL). The organic layer was dried (MgSO₄) and concentrated to give 65 mg of crude 19b and 21b. Flash chromatography on MeOH-deactivated silica gel (4:1 to 2:1 hexanes/EtOAc) gave 5.9 mg (56%) of an inseparable 7:3 mixture of 19b and 21b as a pale yellow gum: IR (CDCl₃) 2933, 1612, 1434, 744; HRMS (EI) calcd C₁₄H₂₀N₂O₂ (M⁺) 248.1525, found 248.1533.

NMR data for **19b** were determined from the mixture: ¹H NMR 6.94–6.92 (m, 1), 6.56–6.54 (m, 1), 6.25–6.21 (m, 1), 4.31–4.23 (m, 1), 3.70 (dd, 1, *J* = 6.1, 4.3), 3.48 (s, 3), 3.03 (dd, 1, *J* = 16.4, 6.1), 2.97 (dd, 1, *J* = 16.4, 4.3), 2.08–1.98 (m, 1), 1.81–1.32 (m, 5), 1.19 (d, 3, *J* = 6.1), the pyrrole NH was not observed; ¹³C NMR 163.1, (127.8 or 127.1), (122.5 or 122.2), (114.2 or 113.6), (109.9 or 109.8), 100.7, 85.3, 68.4, 58.8, 38.6, 34.3, 33.3, 22.3, 20.3.

NMR data for **21b** were determined from the mixture: ¹H NMR 6.92–6.90 (m, 1), 6.56–6.54 (m, 1), 6.25–6.21 (m, 1), 4.07 (dd, 1, J = 6.1, 3.0), 3.81-3.73 (m, 1), 3.38 (s, 3), 3.24 (dd, 1, J = 17.0, 6.1), 2.85 (dd, 1, J = 17.0, 3.0), 2.08-1.98 (m, 1), 1.81-1.32 (m, 5), 1.21 (d, 3, J = 6.1), the pyrrole NH was not observed; ¹³C NMR 164.4, (127.8 or 127.1), (122.5 or 122.2), (114.2 or 113.6), (109.9 or 109.8), 105.0, 83.1, 70.1, 57.5, 39.4, 32.3, 29.1, 22.4, 20.7.

3-Methoxy-α-methyl-5-phenyl-1H-pyrrole-2-butanol (17a). A solution of dimethyl ether **16a** (64 mg, 0.25 mmol) in 10 mL of THF was treated with HF (1.35 M in THF, 3.7 mL, prepared by addition of 1 mL of 48% aqueous HF to 15 mL of THF). The mixture was stirred at 0 °C for 1.5 h. The mixture was diluted with EtOAc, washed with NaHCO₃ (25 mL) and brine (10 mL), dried (Na₂SO₄), and concentrated. Flash chromatography of the residue on MeOH-deactivated silica gel (3:1 hexanes/EtOAc) gave 26 mg (65%) of **17a** as a pale yellow gum.

(\pm)-(5*R*,7*R*)- and (\pm)-(5*S*,7*R*)-4-Methoxy-7-methyl-2-phenyl-6-oxa-1-azaspiro[4,5]deca-1,3-diene (24 and 25). A solution of pyrrole 17a (260 mg, 1 mmol) in 12 mL of acetone was treated with activated MnO₂ (435 mg, 5 mmol) at 25 °C. The mixture was stirred for 20 min and filtered through a pad of Celite. The filtrate was concentrated to give 247 mg of crude product. Flash chromatography of the residue on silica gel (10:1 to 4:1 hexanes/EtOAc) gave 56 mg (14%) of 24 as a yellow gum followed by 90 mg (35%) of 25 as a yellow gum.

Data for 24: ¹H NMR 7.91 (d, 2, J = 7.1), 7.45–7.39 (m, 3), 5.64 (s, 1), 4.45 (dqd, 1, J = 3.0, 6.4, 12.8, H-7), 3.89 (s, 3), 2.22 (ddddd, 1, J = 4.0, 4.0, 12.8, 12.8, 12.8, H-9ax), 1.92 (ddd, 1, J = 3.0, 12.8, 12.8, H-10ax), 1.85 (br d, 1, J = 12.8, H-9eq), 1.76 (br d, 1, J = 12.8, H-8eq), 1.44 (dddd, 1, J = 3.0, 12.8, 12.8, 12.8, 12.8, H-8ax), 1.35 (br d, 1, J = 12.8, H-10eq), 1.21 (d, 3, J = 6.1); ¹³C NMR 182.0, 171.4, 134.9, 130.6, 128.4 (2 C), 127.5 (2 C), 99.7, 92.9, 70.0, 59.0, 32.6, 31.2, 22.1, 21.2; IR (neat) 2934, 1631; HRMS (QTOF) calcd for C₁₆H₂₀NO₂ (MH⁺) 258.1494, found 258.1485.

Data for **25**: ¹H NMR 7.94 (d, 2, J = 7.1), 7.44–7.38 (m, 3), 5.60 (s, 1), 4.43 (dqd, 1, J = 3.0, 6.4, 12.8, H-7), 3.87 (s, 3), 2.15 (br dd, 1, J = 12.8, 12.8, H-10ax), 2.10 (ddddd, 1, J = 3.0, 3.0, 12.8, 12.8, 12.8, H-9ax), 1.84 (ddd, 1, J = 3.0, 3.0, 12.8, H-10eq), 1.72–1.64 (m, 2, H-8eq, H-9eq), 1.44 (dddd, J = 3.0, 12.8, 12.8, 12.8, H-8ax), 1.25 (d, 3, J = 6.1);¹³C NMR 185.6, 171.5, 134.1, 130.8, 128.3 (2 C), 127.7 (2 C), 98.9, 92.2, 69.0, 58.9, 32.0, 30.6, 22.3, 18.5; IR (neat) 2954, 1616; HRMS (QTOF) calcd for C₁₆H₂₀NO₂ (MH⁺) 258.1494, found 258.1496.

(4*S*,5*R*,7*R*)- and (4*R*,5*R*,7*R*)-*rel*-4-Methoxy-7-methyl-2-phenyl-6-oxa-1-azaspiro[4.5]dec-1-ene (18a and 19a). A solution of 24 (13 mg, 0.19 mmol) in 5 mL of MeOH was treated with 5% Pd/ BaSO₄ (20 mg), and the suspension was stirred at 25 °C under H₂ (1 atm) for 3 h. The mixture was then diluted with EtOAc and filtered through a pad of Celite. The filtrate was concentrated to afford 11 mg of crude product. Flash chromatography of the residue on MeOHdeactivated silica gel (5:1 to 2:1 hexanes/EtOAc) gave 5 mg (35%) of 18a as a colorless gum, followed by 3 mg (26%) of 19a as a colorless gum, and then 2 mg (15%) 17a as a pale yellow gum.

(±)-(4S,55,7R)- and (±)-(4R,55,7R)-4-Methoxy-7-methyl-2phenyl-6-oxa-1-azaspiro[4.5]dec-1-ene (20a and 21a). A solution of 25 (21 mg, 0.19 mmol) in 6 mL of MeOH was treated with 5% Pd on BaSO₄ (20 mg), and the suspension was stirred at 25 °C under H₂ (1 atm) for 3 h. The mixture was then diluted with EtOAc and filtered through a pad of Celite. The filtrate was concentrated to afford 46 mg crude product. Flash chromatography of the residue on MeOH-deactivated silica gel (5:1 to 2:1 hexanes/ EtOAc) gave 4 mg (20%) of 21a as a colorless gum, followed by 8 mg (40%) of 20a as a colorless gum, and then 3 mg (15%) 17a as a pale yellow gum.

3-lodo-6-methyl-5,6-dihydropyan-2-one (11). A solution of parascorbic acid $(26)^{23}$ (740 mg, 6.59 mmol) in 12 mL of 1:1 ether/ pyridine was treated with iodine (5.02 g, 19.8 mmol), and the mixture was stirred for 8 h. The reaction was diluted with ether and washed with saturated aqueous Na₂SO₃ (30 mL), saturated aqueous CuSO₄ (3 × 20 mL), and brine (20 mL). The organic layer was dried (Na₂SO₄)

and concentrated. Flash chromatography of the residue on silica gel (6:1 hexanes/EtOAc) gave 1.19 g (76%) of **11** as a pale yellow solid: mp 61–64 °C; ¹H NMR 7.53 (dd, 1, *J* = 3.0, 6.1), 4.72–4.63 (m, 1), 2.47–2.33 (m, 2), 1.44 (d, 3, *J* = 6.1); ¹³C NMR 160.4, 153.7, 89.3, 75.1, 35.0, 20.4; HRMS (QTOF) calcd for $C_6H_8O_2I$ (MH⁺) 238.9569, found 238.9560. The ¹H NMR and ¹³C NMR data are identical to those previously reported for the (*R*) enantiomer.⁴²

3-Phenyl-6-methyl-5,6-dihydro-2*H***-pyran-2-one (27).** A resealable tube was filled with iodolactone **26** (480 mg, 2.0 mmol), phenylboronic acid (726 mg, 6.0 mmol), $Pd(OAc)_2$ (22 mg, 0.1 mmol), SPhos (41 mg, 0.1 mmol), and K_3PO_4 (1.5 g, 7.0 mmol). Degassed *n*-butanol (5 mL) was added, and the mixture was stirred at 80 °C for 6 h. The mixture was diluted with EtOAc and filtered. The filtrate was washed with brine (15 mL), dried (Na₂SO₄), and concentrated. Flash chromatography of the residue on silica gel (8:1 hexanes/EtOAc) gave 322 mg (85%) of **27** as a yellow solid: mp 89–90 °C; ¹H NMR 7.45 (d, 2, *J* = 7.0), 7.40–7.30 (m, 3), 6.94 (dd, 1, *J* = 3.0, 5.9), 4.71–4.61 (m, 1), 2.58–2.43 (m, 2), 1.49 (d, 3, *J* = 6.3); ¹³C NMR 164.5, 140.7, 135.5, 133.1, 128.3 (2 C), 128.2 (3 C), 74.3, 31.8, 20.7; IR (neat) 2976, 1706; HRMS (QTOF) calcd for $C_{12}H_{13}O_2$ (MH⁺) 189.0916, found 189.0915.

(±)-(2S,3S,5S)-5-Hydroxy-2-phenyl-4-propyl-N-methoxy-Nmethyl-hexanamide (31a). A solution of ZnBr₂ (430 mg, 1.83 mmol) in 12 mL of THF was treated with allylmagnesium bromide (1.1 M in THF, 3.33 mL, 3.66 mmol) at 0 °C under nitrogen. The mixture was stirred for 30 min at 0 $^\circ$ C and cooled to -78 $^\circ$ C. A mixture of unsaturated lactone 27 (115 mg, 0.61 mmol) and TMSCl (0.47 mL, 3.66 mmol) in 4 mL of THF was added dropwise. The reaction was stirred at -78 °C for 3 h. Aqueous NH₄Cl solution was added, and the mixture was extracted with EtOAc (3×15 mL). The combined organic layers were washed with brine (15 mL), dried (Na_2SO_4) , and concentrated. Flash chromatography of the residue on silica gel (10:1 hexanes/EtOAc) gave 377 mg (72%) of a 1:3 mixture of 28a and 29a as a pale yellow gum. A 1:3 mixture of 28a and 29a in CDCl₃ was treated with DMAP. The percentage of 28a in the mixture was determined as a function of time by ¹H NMR spectroscopy: initial, 25%; 3 h, 50%; 6 h, 75%; 12 h, 85%. The spectrum did not change at longer times. A similar reaction on a 6:5 mixture also gave a 6:1 mixture of 28a and 29a.

Data of **28a** were determined from a 6:1 mixture of **28a** and **29a**: ¹H NMR 7.36 (t, 2, *J* = 7.6), 7.29 (t, 1, *J* = 7.6), 7.19 (d, 2, *J* = 7.6), 5.72–5.62 (m, 1), 5.09 (d, 1, *J* = 10.1), 5.03 (d, 1, *J* = 17.1), 4.66–4.56 (m, 1), 3.46 (d, 1, *J* = 9.8), 2.34–2.24 (m, 1), 2.16 (ddd, 1, *J* = 4.8, 4.8, 14.0), 1.93 (ddd, 1, *J* = 8.4, 8.4, 14.0), 1.90–1.83 (m, 2), 1.43 (d, 3, *J* = 6.3). A 1D NOESY experiment with irradiation of CHPh at δ 3.46 showed NOEs to the protons at δ 7.19 (ortho phenyl), δ 4.66–4.56 (CHMe), δ 2.34–2.24, and δ 1.93.

Data of **29a** were determined from a 1:3 mixture of **28a** and **29a**: ¹H NMR 7.36–7.24 (m, 3), 7.18 (d, 2, J = 7.6), 5.62–5.50 (m, 1), 5.10–4.98 (m, 2), 4.82–4.73 (m, 1), 3.94 (d, 1, J = 5.6), 2.31–2.22 (m, 1), 2.10–2.02 (m, 2), 1.89–1.80 (m, 1), 1.73 (ddd, 1, J = 3.9, 9.2, 14.0). 1.42 (d, 3, J = 6.1).

A 6:1 mixture of **28a** and **29a** (375 mg, 1.6 mmol) was dissolved in 6 mL of MeOH and treated with a wet slurry of Raney Ni (~30 mg). The suspension was stirred at 25 °C under H₂ (1 atm) for 0.5 h and filtered through a pad of Celite. The filtrate was concentrated to give 341 mg (91%) of a 6:1 mixture of **28b** and **29b** that was used without further purification.

Data of **28b** were determined from the mixture: ¹H NMR 7.37–7.16 (m, 5), 4.66–4.58 (m, 1), 3.40 (d, 1, J = 9.4), 2.20–1.90 (m, 1), 1.89(ddd, 1, J = 8.0, 10.0, 14.4), 1.76(ddd, 1, J = 4.0, 4.0, 14.4), 1.42 (d, 3, J = 6.3), 1.40–1.14 (m, 4), 0.81 (t, 3, J = 7.0); IR (neat) 2929, 1735, 1188.

Partial data of **29b** were determined from the mixture: ¹H NMR 7.37–7.16 (m, 5), 4.83–4.62 (m, 1), 3.89 (d, 1, *J* = 3.9), 2.05 (ddd, 1, *J* = 4.8, 6.8, 11.6).

A 6:1 mixture of lactones **28b** and **29b** (175 mg, 0.75 mmol) and NH(OMe)Me·HCl (300 mg, 3.05 mmol) in 8 mL of THF was treated with *i*-PrMgCl (1.3 M in THF, 6.40 mL) at -20 °C. The reaction was warmed to 0 °C in 30 min and stirred at 0 °C for 2.5 h. Aqueous

 NH_4Cl solution was added, and the mixture was extracted with EtOAc (3 × 15 mL). The organic layers were washed with brine (10 mL), dried (Na_2SO_4), and concentrated. Flash chromatography of the residue on silica gel (3:1 hexanes/EtOAc) gave 29 mg (13%) of **31b** as a colorless gum, followed by 173 mg (78%) of **31a** as a colorless gum.

Data for **31a**: ¹H NMR 7.34–7.21 (m, 5), 3.91 (1, d, J = 10.5), 3.78–3.68 (m, 1), 3.56 (s, 3), 3.15 (s, 3), 2.86 (s, 1, OH), 2.50–2.41 (m, 1), 1.66 (ddd, 1, J = 5.2, 8.8, 14.0), 1.37 (ddd, 1, J = 4.4, 7.6, 14.0), 1.30–1.18 (m, 1), 1.20 (d, 3, J = 6.2), 1.20–1.02 (m, 2), 0.98–0.88 (m, 1), 0.69 (t, 3, J = 7.2); ¹³C NMR 175.5, 138.7, 128.9 (2 C), 128.4 (2 C), 127.0, 64.5, 61.3, 51.8, 43.5, 36.3, 33.9, 32.4, 23.4, 18.7, 14.2; IR (neat) 2931, 1649; HRMS (QTOF) calcd for C₁₇H₂₈NO₃ (MH⁺) 294.2069, found 294.2065.

Data for **31b**: ¹H NMR 7.34 (d, 2, J = 7.2), 7.28 (t, 2, J = 7.2), 7.22 (t, 1, J = 7.2), 4.03 (br d, 1, J = 8.2), 3.60–3.53 (m, 1), 3.53 (s, 3), 3.15 (s, 3), 2.36–2.26 (m, 1), 1.62 (s, 1, OH), 1.42–1.34 (m, 5), 1.32 (ddd, 1, J = 6.8, 6.8, 14.0), 1.21 (ddd, 1, J = 6.0, 6.0, 14.0), 0.97 (d, 3, J = 5.9), 0.91 (t, 3, J = 6.8); IR (neat) 2959, 1642, 1377.

(+)-(2S,3S,5S)-5-Triethylsilyloxy-2-phenyl-4-propyl-N-methoxy-N-methyl-hexanamide (32). A solution of alcohol 31a (87 mg, 0.29 mmol) in THF was treated with TESCI (162 μ L, 0.43 mmol), Et₃N (181 µL, 0.58 mmol), and DMAP (4 mg, 0.03 mmol). The mixture was stirred at 25 °C for 3 h. The reaction was then diluted with Et₂O (10 mL) and washed with brine. The organic layers were dried (MgSO₄) and concentrated. Flash chromatography of the residue on silica gel (10:1 hexanes/EtOAc) gave 102 mg (85%) of 32 as a colorless gum: ¹H NMR 7.35 (d, 2, J = 7.2), 7.28 (t, 2, J = 7.2), 7.21 (t, 1, J = 7.2), 3.97 (br d, 1, J = 9.4), 3.91-3.83 (m, 1), 3.55 (s, 3), 3.12 (s, 3), 2.32-2.22 (m, 1), 1.56 (ddd, 1, J = 5.2, 9.2, 13.6), 1.43 (ddd, 1, J = 3.4, 8.0, 13.6), 1.28–1.10 (m, 4), 1.18 (d, 3, J = 5.9), 0.97 (t, 9, J = 8.0), 0.71 (t, 3, J = 7.1), 0.61 (q, 6, J = 8.0); ¹³C NMR 174.4, 138.5, 129.0 (2 C), 128.2 (2 C), 126.8, 67.2, 61.3, 51.0, 42.0, 37.8, 32.1 (2 C), 23.3, 17.9, 14.4, 6.8 (3 C), 4.8 (3 C); IR (neat) 2956, 1661, 1264; HRMS (QTOF) calcd for C₂₃H₄₁NO₃NaSi (MNa⁺) 430.2753, found 430.2756.

(±)-(4S,5S,7S)-4-Phenyl-5-propyl-7-triethylsilyloxy-1-octen-3-one (33). A solution of Weinreb amide 32 (81 mg, 0.2 mmol) in 6 mL of THF was treated with vinylmagnesium bromide (0.7 M, 0.43 mL) slowly at 25 °C. The reaction was stirred for 1.5 h, and aqueous NH4Cl (5 mL) solution was added. The mixture was extracted with EtOAc (3 \times 10 mL). The organic layers were washed with brine (10 mL), dried (Na_2SO_4) , and concentrated. Flash chromatography of the residue on silica gel (10:1 hexanes/EtOAc) gave 53 mg (72%) of 33 as a colorless gum: ¹H NMR 7.32-7.09 (m, 5), 6.34 (dd, 1, J = 10.1, 17.3), 6.24 (d, 1, J = 17.3), 5.64 (d, 1, J = 10.1), 3.94 (d, 1, J = 9.4), 3.94-3.86 (m, 1), 2.41-2.31 (m, 1), 1.55 (ddd, 1, J = 6.4, 6.4, 13.6), 1.43 (ddd, 1, J = 4.4, 6.8, 13.6), 1.30-1.17 (m, 4), 1.17 (d, 3, J = 6.3), 0.96 (t, 9, J = 8.0), 0.73 (t, 3, J = 7.1), 0.61 (q, 6, J = 8.0); ¹³C NMR 199.8, 137.2, 136.2, 129.3 (2 C), 128.6 (2 C), 127.9, 127.1, 67.0, 60.3, 41.6, 36.5, 32.4, 23.6, 18.3, 14.4, 6.9 (3 C), 5.0 (3 C); IR (neat) 2955, 1699, 1676; HRMS (QTOF) calcd for C23H38O2NaSi (MNa⁺) 397.2539, found 397.2545.

(±)-(25,35,55)-1-[(5R)-4,5-Dihydro-3-[[2-(trimethylsilyl)ethoxy]methyl]-1H-pyrrole-5-isoxazolyl]-2-phenyl-3-propyl-5triethylsilyloxy-1-hexanone (34) and (±)-(25,35,55)-1-[(55)-4,5-Dihydro-3-[[2-(trimethylsilyl)ethoxy]methyl]-1H-pyrrole-5-isoxazolyl]-2-phenyl-3-propyl-5-triethylsilyloxy-1-hexanone (35). A mixture of N-SEM-pyrrole-2-carboxaldehyde oxime (122 mg, 0.51 mmol) and enone 33 (96 mg, 0.26 mmol) in CH₂Cl₂ (15 mL) was treated with bleach (5.25% aqueous NaOCl, 1.12 mL, 58 mg of NaOCl, 0.77 mmol) and Et₃N (13 µL, 0.3 mmol) at 0 °C. The resulting mixture was warmed to 25 °C and stirred for 3 h. The reaction was then diluted with CH2Cl2, washed with brine, dried (Na_2SO_4) , and concentrated. Flash chromatography of the residue on MeOH-deactivated silica gel (18:1 hexanes/EtOAc) gave 30 mg (20%) of pure 34 as a colorless gum, followed by a 4:1 mixture of 34 and 35 (64 mg, 41% yield) as a colorless gum. Further chromatography of this fraction gave a 1:1 mixture of 34 and 35 (24 mg).

Data for 34: ¹H NMR 7.32–7.22 (m, 5), 6.97–6.94 (m, 1), 6.40– 6.37 (m, 1), 6.20–6.17 (m, 1), 5.72 (d, 1, J = 10.2), 5.49 (d, 1, J = 10.2), 4.77 (dd, 1, J = 6.7, 11.5), 4.28 (d, 1, J = 10.2), 3.81 (tq, 1, J = 5.8, 5.8), 3.63 (dd, 1, J = 6.7, 16.8), 3.52 (t, 2, J = 7.8), 3.23 (dd, 1, J = 11.5, 16.8), 2.34–2.25 (m, 1), 1.51 (ddd, 1, J = 5.6, 8.4, 14.0), 1.25–1.12 (m, 4), 1.10 (d, 3, J = 5.8), 0.98–0.90 (m, 1), 0.93 (t, 9, J = 7.8), 0.93 (t, 2, J = 7.8), 0.67 (t, 3, J = 7.0), 0.55 (q, 6, J = 7.8), -0.06 (s, 9); ¹³C NMR 207.2, 150.3, 136.5, 129.5 (2 C), 128.8 (2 C), 127.4, 127.2, 121.4, 115.9, 109.3, 91.5, 81.0, 77.4, 67.2, 65.7, 58.5, 42.0, 37.8, 36.1, 32.2, 23.5, 17.8, 14.5, 6.9 (3 C), 4.9 (3 C), -1.4 (3 C); IR (neat) 2956, 1720, 1358; HRMS (QTOF) calcd for $C_{34}H_{57}N_2O_4Si_2$ (MH⁺) 613.3857, found 613.3860.

Partial data for **35** were obtained from a 1:1 mixture of **34** and **35**: ¹H NMR 7.34–7.12 (m, 5), 6.93–6.92 (m, 1), 6.17–6.15 (m, 1), 6.14–6.12 (m, 1), 5.57 (d, 1, J = 10.2), 5.41 (d, 1, J = 10.2), 4.86 (dd, 1, J = 6.4, 12.0), 4.33 (d, 1, J = 10.2), 3.92–3.84 (m, 1), 3.48 (t, 2, J =7.8), 3.40 (dd, 1, J = 12.0, 16.8), 3.16 (dd, 1, J = 6.4, 16.8), 2.34–2.25 (m, 1), 1.64–1.55 (m, 1), 1.35–1.12 (m, 4), 1.17 (d, 3, J = 5.8), 0.90 (t, 9, J = 7.8), 0.63 (q, 6, J = 7.8), -0.06 (s, 9).

(\pm)-(4*R*,5*R*,75,95,105)-4-Methoxy-7-methyl-9-propyl-10-phenyl-2-(1-[[2-(trimethylsilyl)ethoxy]methyl]-1*H*-pyrrol-2-yl)-6oxa-1-azaspiro[4.5]dec-1-ene (36). A solution of pure isoxazoline 34 (29 mg, 47 μ mol) in 6 mL of MeOH was treated with a wet slurry of Raney nickel 2800 (~20 mg), and the suspension was stirred at 25 °C under H₂ (1 atm) for about 30 min. The mixture was then diluted with EtOAc and filtered. The filtrate was washed with brine (3 × 5 mL), dried (Na₂SO₄), and concentrated to give 27 mg of crude hydroxy hemi-iminal.

A solution of crude hydroxy hemi-iminal in THF (1 mL) was added dropwise to a suspension of NaH (60% in mineral oil, 15 mg, 0.37 mmol) in THF (5 mL) at 0 °C. The mixture was stirred at 0 °C for 10 min, and MeI (23 μ L, 0.37 mmol) was added dropwise by syringe over 2 min. The resulting mixture was warmed to 25 °C and stirred for 4 h. The reaction was quenched with saturated aqueous NH₄Cl (3 mL), and the aqueous layer was extracted with EtOAc. The combined organic layers were dried (Na₂SO₄) and concentrated to give 23 mg of crude dimethyl ether. Flash chromatography on silica gel (25:1 hexanes/EtOAc) gave 13 mg (45% for two steps) of a single isomer of the dimethyl ether as a pale yellow gum: ¹H NMR 7.16–7.10 (m, 5), 7.06–7.03 (m, 1), 6.50–6.46 (m, 1), 6.21–6.18 (m, 1), 6.01 (d, 1, J = 9.8), 5.77 (d, 1, J = 9.8), 3.98 (tq, 1, J = 5.9, 5.9), 3.64 (dd, 1, J = 3.1, 7.9), 3.50 (t, 2, *J* = 8.2), 3.43 (s, 3), 3.34 (s, 3), 3.03 (d, 1, *J* = 4.3), 2.64 (dd, 1, J = 3.1, 17.4), 2.58-2.48 (m, 1), 2.40 (dd, 1, J = 7.9, 17.4),2.15-2.05 (m, 1), 1.46-1.14 (m, 4), 1.19 (d, 3, J = 5.9), 0.98 (t, 9, J = 5.9)7.8), 0.94-0.80 (m, 6), 0.64 (q, 6, J = 7.8), -0.03 (s, 9); IR (neat) 2955, 1615.

A solution of the dimethyl ether (13 mg, 21 μ mol) in 8 mL of 3:1 CH₃CN/THF was treated with aqueous 2 M HCl (210 μ L, 420 μ mol) at 25 °C. The resulting mixture was stirred at 25 °C for 20 h. Saturated NaHCO₃ (4 mL) was added to bring the pH to 7. The reaction was extracted with EtOAc, and the organic layer was washed with brine, dried (Na₂SO₄), and concentrated to give 12.5 mg of crude 36. Flash chromatography of the residue on MeOH-deactivated silica gel (6:1 hexanes/EtOAc) gave 7.2 mg (70%) of 36 as a colorless gum: $^1\mathrm{H}$ NMR (CDCl₃) 7.25-7.18 (m, 2), 7.16-7.11 (m, 3), 7.02-6.98 (m, 1), 6.37–6.33 (m, 1), 6.27 (d, 1, J = 9.8), 6.15–6.11 (m, 1), 5.71 (d, 1, *J* = 9.8), 4.50–4.41 (m, 1), 3.64–3.53 (m, 3), 3.36 (s, 3), 2.84 (d, 1, *J* = 10.9), 2.66 (dd, 1, J = 5.2, 16.2), 2.62–2.51 (m, 1), 2.06 (dd, 1, J = 7.0, 16.2), 1.99 (ddd, 1, J = 4.0, 4.0, 12.8), 1.73 (ddd, 1, J = 5.6, 9.6, 12.8), 1.47 (d, 3, J = 6.6), 1.40–1.27 (m, 1), 1.18–1.02 (m, 2), 1.02– 0.88 (m, 3), 0.74 (t, 3, J = 7.1), 0.00 (s, 9); ¹H NMR (acetone- d_6) 7.36–7.30 (m, 2), 7.18–7.10 (m, 4), 6.42–6.38 (m, 1), 6.32 (d, 1, J = 9.8), 6.11-6.08 (m, 1), 5.73 (d, 1, J = 9.8), 4.40-4.30 (m, 1), 3.66 (t, 2, *J* = 7.1), 3.56 (dd, 1, *J* = 4.2, 7.0), 3.39 (s, 3), 2.90 (d, 1, *J* = 11.3), 2.62-2.51 (m, 1), 2.50 (dd, 1, J = 4.2, 16.2), 2.12-2.03 (m, 1), 1.90 (dd, 1, J = 7.0, 16.2), 1.73 (ddd, 1, J = 4.8, 7.6, 12.8), 1.40 (d, 3, J = 6.6), 1.42-1.33 (m, 1), 1.20-1.07 (m, 2), 1.07-0.82 (m, 3), 0.74 (t, 3, J = 7.1, 0.00 (s, 9); ¹³C NMR (CDCl₃) 164.3, 140.1, 130.4 (br, 2 C), 127.7 (2 C), 127.3, 127.0, 126.3, 116.9, 108.4, 103.4, 82.1, 77.4, 69.4, 65.7, 58.4, 56.0, 39.6, 36.1, 35.8, 31.8, 23.4, 19.2, 18.1, 14.2, -1.4 (3

C); 13 C NMR (acetone- d_6) 164.8, 141.3, 131.5 (br, 2 C), 129.1 (br, 2 C), 128.6, 128.2, 127.2, 117.9, 109.1, 105.1, 83.2, 78.0, 68.9, 66.0, 58.7, 56.9, 41.5, 37.1, 36.9, 33.3, 23.9, 20.1, 18.7, 14.6, -1.2 (3 C); IR (neat) 2925, 1616, 1084. HRMS (QTOF) calcd for C₂₉H₄₅N₂O₃Si (MH⁺) 497.3199, found 497.3192.

A 2D NOESY experiment in CDCl₃ showed NOEs between CHOMe H-4 at δ 3.64–3.53 and both CHPh H-10 at δ 2.84 and the phenyl protons at δ 7.25–7.18 and between CHPr H-9 at δ 2.62–2.51 and 7-Me at δ 1.47.

(±)-(45,5*R*,75,95,105)-4-Methoxy-7-methyl-9-propyl-10-phenyl-2-(1-[[2-(trimethylsilyl)ethoxy]methyl]-1*H*-pyrrol-2-yl)-6oxa-1-azaspiro[4.5]dec-1-ene (37). A 1:1 mixture of isoxazolines 34 and 35 (24 mg, 39 μ mol) in 5 mL of MeOH was treated with a wet slurry of Raney nickel 2800 (~20 mg), and the suspension was stirred at 25 °C under H₂ (1 atm) for about 30 min. The mixture was then diluted with EtOAc and filtered. The filtrate was washed with brine (3 × 5 mL), dried (Na₂SO₄), and concentrated to give 22 mg of crude hydroxy hemi-iminals.

A solution of crude hydroxy hemi-iminals in THF (1 mL) was added dropwise to a suspension of NaH (60% in mineral oil, 15 mg, 0.37 mmol) in THF (5 mL) at 0 °C. The mixture was stirred at 0 °C for 10 min, and MeI (23 μ L, 0.37 mmol) was added dropwise by syringe over 2 min. The resulting mixture was warmed to 25 °C and stirred for 4 h. The reaction was quenched with saturated aqueous NH₄Cl (3 mL), and the aqueous layer was extracted with EtOAc. The combined organic layers were dried (Na₂SO₄) and concentrated to give 17 mg of crude mixture of dimethyl ethers. Flash chromatography on silica gel (25:1 hexanes/EtOAc) gave 11 mg (45% for two steps) of a 1:1 mixture of two diastereomeric dimethyl ethers as a colorless gum.

Data for the isomer not obtained from 34 were determined from the mixture: ¹H NMR 7.46–7.41 (d, 2, J = 7.0), 7.28–7.19 (m, 3), 7.06–7.03 (m, 1), 6.59–6.56 (m, 1), 6.46 (d, 1, J = 9.8), 6.21–6.18 (m, 1), 5.54 (d, 1, J = 9.8), 3.85 (tq, 1, J = 5.9, 5.9), 3.66 (d, 1, J = 3.0, 7.2), 3.61–3.54 (m, 2), 3.36 (s, 3), 3.08 (dd, 1, J = 7.2, 17.6), 2.96 (s, 3), 2.82 (dd, 1, J = 3.0, 17.6), 2.55 (d, 1, J = 4.8), 2.29–2.20 (m, 1), 2.15–2.05 (m, 1), 1.46–1.14 (m, 4), 1.06 (d, 3, J = 5.9), 0.92–0.80 (m, 6), 0.87 (t, 9, J = 7.8), 0.49 (m, 6, J = 7.8), -0.03 (s, 9).

A 1:1 mixture of dimethyl ethers (22 mg, 36 μ mol) in 8 mL of 3:1 CH₃CN/THF was treated with aqueous 2 M HCl (360 μ L, 720 μ mol) at 25 °C. The resulting mixture was stirred at 25 °C for 20 h. Saturated NaHCO₃ (10 mL) was added to bring the pH to 7. The reaction was extracted with EtOAc, and the organic layer was washed with brine, dried (Na₂SO₄), and concentrated to give 24 mg of a crude mixture of **36** and **37**. Flash chromatography of the residue on MeOH-deactivated silica gel (10:1 hexanes/EtOAc) gave 5.6 mg (32%) of **37** as a colorless gum, followed by 5.0 mg (29%) of **36** as a colorless gum.

Data for 37: ¹H NMR (CDCl₃) 7.23 (d, 2, J = 7.2), 7.07–7.6.98 (m, 3), 6.98–6.96 (m, 1), 6.19 (d, 1, J = 10.0), 6.18–6.15 (m, 1), 6.11– 6.08 (m, 1), 5.70 (d, 1, J = 10.0), 4.44-4.36 (m, 1), 3.80 (dd, 1, J = 8.8, 8.8), 3.66-3.55 (m, 2), 3.42 (s, 3), 3.01 (d, 1, J = 11.6), 2.61 (dd, 1, J = 8.8, 15.6), 2.47-2.36 (m, 1), 1.96 (ddd, 1, J = 4.0, 4.0, 13.6),1.70 (ddd, 1, J = 5.2, 10.0, 13.6), 1.57 (d, 3, J = 6.8), 1.38–1.26 (m, 1), 1.11 (dd, 1, J = 8.8, 15.6), 1.16-0.88 (m, 5), 0.72 (t, 3, J = 7.1), 0.00 (s, 9); ¹H NMR (acetone- d_6) 7.32 (d, 2, J = 7.2), 7.12–7.09 (m, 1), 7.09–6.92 (m, 3), 6.29–6.23 (m, 3), 6.08–6.04 (m, 1), 5.71 (d, 1, J = 10.0), 4.38–4.29 (m, 1), 3.78 (dd, 1, J = 8.8, 8.8), 3.66 (t, 2, J = 7.1), 3.41 (s, 3), 3.00 (d, 1, J = 11.4), 2.69 (dd, 1, J = 8.8, 15.6), 2.55-2.45 (m, 1), 1.97 (ddd, 1, J = 3.6, 3.6, 13.6), 1.67 (ddd, 1, J = 6.4, 10.0, 13.6), 1.56 (d, 3, J = 7.1), 1.41–1.28 (m, 1), 1.10 (dd, 1, J = 8.8, 15.6), 1.20–0.85 (m, 5), 0.72 (t, 3, J = 7.1), 0.00 (s, 9); ¹³C NMR (CDCl₃) 163.0, 140.0, 131.5 (br, 2 C), 127.24, 127.19, 126.7 (2 C), 125.7, 116.8, 108.4, 105.4, 88.3, 69.5, 65.5, 58.3, 52.1, 39.2, 35.7 (2 C), 31.2, 22.9, 19.2, 18.0, 14.1, -1.4 (3 C), one peak is obscured by the CDCl₃ triplet at δ 77.0; ¹³C NMR (acetone-d₆) 163.7, 141.3, 132.6 (br, 2 C), 129.1 (br, 2 C), 128.0, 127.5, 126.5, 117.8, 109.0, 106.3, 89.4, 77.9, 69.9, 65.9, 58.5, 53.2, 40.1, 36.8, 36.7, 32.2, 23.5, 20.0, 18.6, 14.6, -1.2 (3 C); IR (neat) 2928, 1619; HRMS (QTOF) calcd for C₂₉H₄₅N₂O₃Si (MH⁺) 497.3199, found 497.3199.

A 2D NOESY experiment showed NOEs between the OMe at δ 3.42 and CHPh H-10 at δ 3.01 and the phenyl protons at δ 7.23 and between CHPr H-9 at δ 2.47–2.36 and 7-Me at δ 1.57.

2-(Hex-5-enyl)-1H-pyrrole (39). Pyrrole 39 was prepared by Muchowski's procedure.^{3,30} Magnesium ribbon (400 mg, 16.67 mmol) and a small crystal of iodine (~20 mg) were placed in a 100-mL flask. The flask was flushed with nitrogen and was treated with 30 mL of THF. The suspension of magnesium in THF was slowly treated with 4-bromo-1-pentene (1.64 g, 11.0 mmol) and was heated to reflux gently. The resulting solution was refluxed for 2 h, cooled to 25 °C, and cannulated to a solution of 1-(phenylsulfonyl)-2-pyrrolecarboxaldehyde⁴³ (1.95 g, 8.25 mmol) in THF (15 mL) at 0 °C. The mixture was then stirred at 25 °C for 3 h and was quenched with 0.5 M HCl (8 mL). The aqueous layer was extracted with EtOAc, and the combined organic layers were dried (Na₂SO₄) and concentrated. Flash chromatography on silica gel (5:1 hexanes/EtOAc) gave 1.62 g (64%) of α -5-hexen-1-yl-1-(phenylsulfonyl)-1*H*-pyrrole-2-methanol as a pale yellow gum: ¹H NMR 7.78 (d, 2, J = 7.4), 7.62 (t, 1, J = 7.4), 7.51 (t, 2, J = 7.4), 7.30 (dd, 1, J = 1.5, 3.3), 6.32-6.25 (m, 2), 5.79-5.68 (m, 1), 4.98-4.90 (m, 2), 4.81 (t, 1, J = 6.0), 2.74 (br s, 1, OH), 2.06-1.92 (m, 2), 1.89-1.73 (m, 2), 1.54-1.42 (m, 1), 1.42-1.31 (m, 1); ¹³C NMR 139.2, 138.4, 138.2, 133.9, 129.4 (2 C), 126.4 (2 C), 123.5, 114.7, 112.4, 111.7, 65.0, 34.4, 33.2, 25.2; IR (neat) 3554, 1364, 1175; HRMS (QTOF) calcd for C₁₆H₁₉NO₃NaS (MNa⁺) 328.0983, found 328.0980.

A solution of the above alcohol (1.62 g, 5.26 mmol) in CH_2Cl_2 (20 mL) was treated with molecular sieves (4 Å, 2.5 g), *N*-methylmorpholine-*N*-oxide (1.23 g, 10.52 mmol), and tetrapropylammonium perruthenate (0.19 g, 0.53 mmol) at 0 °C. The reaction was stirred at 25 °C for 4 h and filtered through a pad of Celite. The filtrate was concentrated to afford 2.78 g of reaction crude as a black oil. Flash chromatography on silica gel (8:1 hexanes/EtOAc) gave 1.44 g (89%) of 1-(1-(phenylsulfonyl)-1H-pyrrol-2-yl)hex-5-en-1-one as a pale yellow gum: ¹H NMR 7.99 (d, 2, *J* = 7.2), 7.80 (dd, 1, *J* = 3.2, 1.6), 7.59 (t, 1, *J* = 7.2), 7.52 (t, 2, *J* = 7.2), 7.03 (dd, 1, *J* = 3.2, 1.6), 7.34 (t, 1, *J* = 1.6), 5.78–5.66 (m, 1), 5.01–4.92 (m, 2), 2.67 (t, 2, *J* = 7.6), 2.00 (dt, 2, *J* = 7.6, 7.6), 1.69 (tt, 2, *J* = 7.6, 7.6); ¹³C NMR (rotamer) 188.7, 138.9, 137.8, 133.5, 133.4, 130.10 (130.06), 128.7 (2 C), 128.1 (2 C), 123.3, 115.3 (br), 110.4 (110.3), 38.4, 32.9, 23.7; IR (neat) 1672, 1438, 1141; HRMS (QTOF) calcd for $C_{16}H_{18}NO_3S$ (MH⁺) 304.1007, found 304.0999.

A solution of the ketone (1.44 g, 4.75 mmol) in *i*-PrOH (50 mL) was treated with NaBH₄ (1.26 g, 33.3 mmol). The mixture was refluxed for 16 h, cooled to 25 °C, and slowly quenched with water (50 mL) and saturated aqueous NH₄Cl (30 mL). The mixture was extracted with EtOAc (3×20 mL). The combined organic layers were dried (MgSO₄) and concentrated to give 1.06 g of crude **39**. Flash chromatography on silica gel (12:1 hexanes/EtOAc) gave 0.65 g (84%) of **39** as a pale colorless gum with data identical to those previously reported.³¹

2-(Hex-5-enyl)-pyrrole-1-carboxylic Acid *tert***-Butyl Ester (40).** A solution of pyrrole 39^{31} (654 mg, 4.08 mmol) in 8 mL of CH₂Cl₂ was treated with (Boc)₂O (1.24 g, 5.69 mmol), Et₃N (0.82 mL, 5.69 mmol), and DMAP (30 mg, 0.43 mmol). The resulting solution was stirred for 5 h and concentrated. Flash chromatography of the residue on silica gel (25:1 hexanes/EtOAc) gave 811 mg (74%) of 40 as a colorless gum: ¹H NMR 7.19 (dd, 1 *J* = 1.8, 3.3), 6.08 (t, 1, *J* = 3.3), 5.96 (dd, 1, *J* = 1.8, 3.3), 5.82 (ddt, 1, *J* = 10.2, 17.2, 6.7), 5.01 (ddt, 1, *J* = 1.4, 17.2, 1.4), 4.95 (ddt, 1, *J* = 1.4, 10.2, 1.0), 2.85 (t, 2, *J* = 7.6), 2.10 (dt, 2, *J* = 6.7, 7.6), 1.64 (tt, 2, *J* = 7.6, 7.6), 1.59 (s, 9), 1.48 (tt, 2, *J* = 7.6, 7.6); ¹³C NMR (rotamer) 149.5, 138.9 (138.8), 136.3, 120.8 (120.7), 114.3, 110.8 (110.6), 109.9 (109.7), 83.1, 33.6 (br), 28.7, 28.6, 28.3, 28.0 (3 C); IR (neat) 2935, 1742, 1330; HRMS (EI) calcd for C₁₅H₂₃NO₂ (M⁺) 249.1729, found 249.1733.

3-[5-(Hex-5-enyl)-1-*tert***-butoxycarbonylpyrrol-2-yl]-6-meth-yl-5,6-dihydro-2H-pyran-2-one (42).** A solution of 2,2,6,6-tetramethylpiperidine (1.37 mL, 8.13 mmol) in 12 mL of THF was treated with *n*-BuLi (1.6 M in THF, 5.08 mL) dropwise at -78 °C under nitrogen. The solution was stirred for 15 min, warmed to 0 °C for 30 min, and recooled to -78 °C. A solution of pyrrole 40 (1.35 g,

5.40 mmol) in 2 mL of THF was added dropwise, and the reaction mixture was stirred at -78 °C for 2 h. Trimethyl borate (6.3 mL, 27.0 mmol) was added at -78 °C, and the solution was warmed to 25 °C over 2 h and stirred overnight. Aqueous HCl (0.2 N, 45 mL) was added, and the resulted mixture was extracted with EtOAc (3× 20 mL). The combined organic layers were washed with brine (25 mL) and dried (Na₂SO₄). The solution was slowly concentrated until a white solid started to precipitate. Then 5 mL of dry 1,2-dimethoxy-ethane was added, and the solution was slowly concentrated to 2 mL to remove the remaining EtOAc. The solution was used immediately after being degassed.

A resealable tube was filled with iodolactone 26 (400 mg, 1.67 mmol), LiCl (214 mg, 5.00 mmol), Na₂CO₃ (1.06 g, 10.0 mmol), and $Pd(PPh_3)_4$ (155 mg, 0.13 mmol) and was purged with nitrogen. Then 2 mL of the above 1,2-dimethoxyethane solution of boronic acid 41 was added to the mixture, after which 0.3 mL of degassed H₂O was added to the mixture. The tube was sealed and stirred at 80 °C for 8 h. The mixture was diluted with EtOAc and filtered. The filtrate was washed with brine (15 mL), dried (Na₂SO₄), and concentrated. Flash chromatography of the residue on silica gel (8:1 hexanes/EtOAc) gave 330 mg (55%) of 42 as a yellow gum: ¹H NMR 6.74 (dd, 1, J = 3.1, 5.5), 6.06 (d, 1, J = 2.7), 5.88 (d, 1, J = 2.7), 5.81 (ddd, 1, J = 6.6, 10.2, 17.2), 5.00 (br d, 1, J = 17.2), 4.94 (br d, 1, J = 10.2), 4.76-4.67 (m, 1), 2.85–2.72 (m, 2), 2.48–2.35 (m, 2), 2.08 (m, 2), 1.65–1.40 (m, 4), 1.55 (s, 9), 1.47 (d, 3, J = 6.1); ¹³C NMR 164.3, 150.1, 138.8, 137.4, 136.9, 129.3, 128.6, 114.4, 112.6, 109.3, 84.1, 74.2, 33.6, 31.4, 29.3, 28.7, 28.5, 27.9 (3 C), 20.7; IR (neat) 1730 (br), 1367, 1216; HRMS (QTOF) calcd for $C_{21}H_{30}NO_4$ (MH⁺) 360.2175, found 360.2177

(±)-(35,45,65)-3-[5-(Hex-5-enyl)-1*H*-pyrrol-2-yl]-4-(2-propen-1-yl)-6-methyl-3,4,5,6-tetrahydro-2*H*-pyran-2-one (45). A solution of ZnBr₂ (1.00 g, 4.44 mmol) in 20 mL of THF was treated with allylmagnesium bromide (1.7 M in THF, 5.22 mL) at 0 °C under nitrogen. The mixture was stirred for 30 min at 0 °C and cooled to -78 °C. A mixture of unsaturated lactone 42 (400 mg, 1.11 mmol) and TMSCl (1.13 mL, 4.44 mmol) in 4 mL of THF was added dropwise. The reaction was stirred at -78 °C for 3 h. Aqueous NH₄Cl solution was added, and the mixture was extracted with EtOAc (3 × 15 mL). The combined organic layers were washed with brine (15 mL), dried (Na₂SO₄), and concentrated. Flash chromatography of the residue on silica gel (10:1 hexanes/EtOAc) gave 249 mg (56%) of a 5:4 mixture of *cis*-addition product 43 and Boc-migration product 44 as a pale yellow gum

Partial data of **43** were determined from the mixture: ¹H NMR 6.04–6.02 (m, 1), 5.90–5.86 (m, 1), 5.86–5.72 (m, 1), 5.65–5.52 (m, 1), 5.20–4.82 (m, 4), 4.72 (d, 1, J = 4.9), 4.72–4.64 (m, 1), 2.80–2.70 (m, 2), 2.48–2.30 (m, 2), 2.20–1.70 (m, 5), 1.59 (s, 9), 1.62–1.40 (m, 4), 1.39 (d, 3, J = 6.1).

Partial data of 44 were determined from the mixture: ¹H NMR 5.98–5.95 (m, 1), 5.90–5.86 (m, 1), 5.86–5.65 (m, 2), 5.20–4.82 (m, 4), 4,60–4.52 (m, 1), 2.80–2.70 (m, 2), 2.48–2.30 (m, 1), 2.20–1.70 (m, 6), 1.59 (s, 9), 1.62–1.40 (m, 4), 1.41 (d, 3, J = 6.1).

A solution of the mixture of 43 and 44 (270 mg, 0.67 mmol) in CH₂Cl₂ was treated with 2,6-lutidine (0.43 mL, 4.05 mmol) and TMSOTf (0.6 mL, 2.7 mmol) at 0 °C. The reaction was stirred at 25 °C for 8 h, diluted with CH₂Cl₂, and treated with AcOH (0.5 mL). The mixture was washed with water (10 mL), NaHCO₃ (5 mL), and brine (3 × 10 mL). The organic phase was dried (Na₂SO₄) and concentrated. Flash chromatography of the residue on silica gel (6:1 hexanes/EtOAc) gave 182 mg (91%) of a 19:1 equilibrium mixture of **45** and the *cis* isomer of **45** as a pale yellow gum.

Data of **45** were determined from the mixture: ¹H NMR 8.39 (br s, 1, NH), 5.92 (t, 1, J = 2.7), 5.79 (t, 1, J = 2.7), 5.86–5.69 (m, 2), 5.11 (d, 1, J = 10.0), 5.09 (d, 1, J = 17.2), 5.00 (d, 1, J = 17.2), 4.94 (d, 1, J = 10.4), 4.64–4.58 (m, 1), 3.57 (d, 1, J = 6.7), 2.56 (t, 2, J = 7.6), 2.44–2.30 (m, 2), 2.17–2.10 (m, 1), 2.07 (dt, 2, J = 7.0, 7.0), 1.89 (ddd, 1, J = 6.4, 10.0, 14.4), 1.80 (ddd, 1, J = 3.6, 3.6, 14.4), 1.62 (tt, 2, J = 7.6, 7.6), 1.45 (tt, 2, J = 7.6, 7.6), 1.37 (d, 3, J = 6.4); ¹³C NMR 173.1, 138.8, 134.9, 133.4, 124.2, 118.0, 114.5, 106.5, 104.6, 73.7, 43.6,

38.4, 33.6, 33.5, 32.9, 28.9, 28.6, 27.6, 21.3; IR (neat) 3365, 1704; HRMS (QTOF) calcd for $C_{19}H_{28}NO_2$ (MH⁺) 302.2120, found 302.2114. A 2D NOESY experiment showed NOEs between H-3 at δ 3.57 and H-6 at δ 4.64–4.58.

Partial data for the *cis* isomer of **45** were determined from the mixture: ¹H NMR 4.09 (d, 1, J = 6.8).

(±)-(35,4a5,15a5)-3-Methyl-3,4,4a,5,6,7,8,9,10,11-decahydro-12,15-epiminocycloundeca[c]pyran-1(15aH)-one (47b). A solution of diene 45 (40 mg, 0.13 mmol) in 250 mL of degassed CH_2Cl_2 was slowly treated with a solution of Grubbs II catalyst (8 mg, 72 µmol) in 5 mL of degassed CH_2Cl_2 at reflux under nitrogen. The solution was stirred at reflux for 8 h, and another solution of Grubbs II catalyst (8 mg, 72 µmol) in 5 mL of CH_2Cl_2 was slowly added. The solution was stirred at reflux for another 8 h. The reaction was cooled to 25 °C, and five drops of DMSO were added. The solution was stirred overnight and concentrated. Flash chromatography of the residue on silica gel (6:1 hexanes/EtOAc) gave 16 mg (41%) of **46** as a brown solid, which was used directly for the next step.

A solution of alkene **46** (48 mg, 0.18 mmol, from three runs of the previous reaction) in 5 mL of MeOH was treated with a wet slurry of Raney nickel 2800 (~15 mg), and the suspension was stirred at 25 °C under H₂ (1 atm) for 25 min. The mixture was then diluted with EtOAc and filtered through a pad of Celite. The filtrate was concentrated and filtered again through a pad of silica gel to give 43 mg (90%) of **47b** as a white solid: mp 165 °C (decomposed); ¹H NMR 8.23 (br s, 1, NH), 5.92 (m, 1), 5.77 (m, 1), 4.64–4.54 (m, 1), 3.47 (d, 1, *J* = 12.1), 2.55 (ddd, 1, *J* = 4.4, 4.4, 14.4), 2.44 (ddd, 1, *J* = 3.8, 10.8, 14.4), 2.20–2.09 (m, 1), 2.04 (ddd, 1, *J* = 10.0, 10.0, 14.0), 1.67 (ddd, 1, *J* = 4.0, 4.0, 14.0), 1.64–1.44 (m, 4), 1.39 (d, 3, *J* = 6.4), 1.37–1.02 (m, 5), 0.91–0.72 (m, 2), 0.48–0.35 (m, 1); ¹³C NMR 175.0, 134.0, 124.4, 110.2, 105.7, 73.1, 45.5, 38.3, 33.1, 32.5, 28.1, 26.8, 25.5, 24.6, 24.2, 24.1, 20.9; IR (neat) 3343, 2924, 1725, 1211; HRMS (QTOF) calcd for C₁₇H₂₆NO₂ (MH⁺) 276.1964, found 276.1969.

(±)-(2S,3S)-3-((S)-2-Hydroxy)propyl)-N-methoxy-N-methyl-14-azabicyclo[9.2.1]tetradeca-1(13),11-diene-2-carboxamide (48b). A mixture of lactone 47b (43 mg, 0.16 mmol) and NH(OMe)Me·HCl (64 mg, 0.65 mmol) in 8 mL of THF was treated with *i*-PrMgCl (1.3 M in THF, 1.37 mL) at -20 °C. The reaction was warmed to 0 °C in 30 min and stirred at 0 °C for 2.5 h. Aqueous NH₄Cl solution was added, and the mixture was extracted with EtOAc (3 \times 15 mL). The combined organic layers were washed with brine (10 mL), dried (Na₂SO₄), and concentrated. Flash chromatography of the residue on silica gel (3:1 hexanes/EtOAc) gave 43 mg (86%) of 48b as a colorless gum: ¹H NMR 8.43 (br s, 1, NH), 5.86 (m, 1), 5.74 (m, 1), 3.91 (d, 1, J = 11.0), 3.75 (s, 3), 3.74–3.65 (m, 1), 3.19 (s, 3), 2.72 (br s, 1, OH), 2.65 (ddd, 1, J = 4.0, 4.0, 14.4), 2.49 (ddd, 1, J =3.2, 11.2, 14.4), 2.28–2.17 (m, 1), 1.75–1.20 (m, 9), 1.18 (d, 3, J = 6.1), 1.18–1.04 (m, 2), 1.04–0.74 (m, 3), 0.47–0.35 (m, 1); ^{13}C NMR 175.5, 133.7, 127.2, 108.5, 105.3, 64.8, 61.6, 46.4, 45.0, 35.1, 32.2, 28.8, 28.2, 27.8, 25.2, 25.0, 24.8, 24.5, 23.3; IR (neat) 2958, 1667; HRMS (QTOF) calcd for $C_{19}H_{33}N_2O_3$ (MH+) 337.2491, found 337.2485.

(+)-(2S,3S)-N-Methoxy-N-methyl-3-((S)-2-((triethylsilyl)oxy)propyl)-14-azabicyclo[9.2.1]tetradeca-1(13),11-diene-2-carboxamide (49b). A solution of alcohol 48b (43 mg, 0.16 mmol) in 3 mL of THF was treated with TESCl (90 μ L, 0.24 mmol), Et₃N (100 μ L, 0.32 mmol), and DMAP (2 mg, 0.02 mmol). The mixture was stirred at 25 °C for 3 h. The reaction was then diluted with Et₂O (10 mL) and washed with brine (10 mL). The organic layer was dried (MgSO₄) and concentrated. Flash chromatography of the residue on silica gel (10:1 hexanes/EtOAc) gave 52 mg (77%) of 49b as a pale yellow gum: ¹H NMR 8.49 (br s, 1, NH), 5.83 (t, 1, *J* = 2.7), 5.74 (t, 1, J = 2.7), 3.88 (tq, 1, J = 6.1, 6.1), 3.85 (d, 1, J = 11.0), 3.71 (s, 3), 3.17 (s, 3), 2.65 (ddd, 1, *J* = 4.6, 4.6, 14.6), 2.46 (ddd, 1, *J* = 3.7, 11.2, 14.6), 2.10-1.98 (m, 1), 1.79-1.44 (m, 3), 1.46-1.20 (m, 7), 1.18 (d, 3, J = 6.1), 1.18–1.02 (m, 2), 0.96 (t, 9, J = 8.0), 0.92–0.72 (m, 2), 0.59 (q, 6, J = 8.0), 0.45-0.35 (m, 1); ¹³C NMR 174.5, 133.2, 127.4, 108.1, 105.6, 66.7, 61.5, 46.9, 45.8, 35.9, 31.9, 29.1, 28.1, 26.6, 25.9, 25.2, 24.9, 24.6, 23.0, 6.9 (3 C), 4.8 (3 C); IR (neat) 2935, 1679, 1268;

HRMS (QTOF) calcd for $\rm C_{25}H_{46}N_2O_3NaSi~(MNa^+)$ 473.3175, found 473.3173.

(±)-1-((25,35)-3-((5)-2-((Triethylsilyl)oxy)propyl)-14azabicyclo[9.2.1]tetradeca-1(13),11-dien-2-yl)prop-2-en-1-one (50b). A solution of vinyl bromide $(24 \,\mu\text{L}, 76 \,\mu\text{mol})$ in 4 mL of ether was treated with *n*-BuLi (1.6 M in THF, 48 μ L) dropwise at -78 °C. Then the solution was transferred to a solution of Weinreb amide 49b (17 mg, 38 μ mol) in 2 mL of ether by cannula at 0 °C. The mixture was stirred at 0 °C for 2 h. Aqueous NH4Cl solution was added and extracted with EtOAc $(3 \times 5 \text{ mL})$. The organic solution was washed with brine (5 mL), dried (Na2SO4), and concentrated. Flash chromatography of the residue on silica gel (16:1 hexanes/EtOAc) gave 11 mg (70%) of 50b as a pale yellow gum (containing 5% of bisvinyl tertiary alcohol): ¹H NMR 8.13 (br s, 1, NH), 6.44 (dd, 1, J = 10.4, 17.6), 6.22 (d, 1, J = 17.6), 5.88 (t, 1, J = 2.8), 5.80 (d, 1, J = 17.6), 5.88 (t, 1, J = 2.8), 5.80 (d, 1, J = 17.6), 5.88 (t, 1, J = 2.8), 5.80 (d, 1, J = 17.6), 5.88 (t, 1, J = 2.8), 5.80 (d, 1, J = 17.6), 5.88 (t, 1, J = 2.8), 5.80 (d, 1, J = 17.6), 5.88 (t, 1, J = 2.8), 5.80 (d, 1, J = 17.6), 5.88 (t, 1, J = 2.8), 5.80 (d, 1, J = 17.6), 5.88 (t, 1, J = 2.8), 5.80 (d, 1, J = 17.6), 5.88 (t, 1, J = 2.8), 5.80 (t, 1, J = 1.6), 5.88 (t 10.4), 5.75 (t, 1, J = 2.8), 3.90 (tq, 1, J = 6.1, 6.1), 3.86 (d, 1, J = 10.2), 2.63 (ddd, 1, J = 4.6, 4.6, 14.6), 2.47 (ddd, 1, J = 3.7, 11.2, 14.6), 2.18-2.10 (m, 1), 1.62–1.50 (m, 2), 1.44–1.24 (m, 7), 1.18 (d, 3, J = 6.1), 1.18–1.02 (m, 2), 0.96 (t, 9, J = 8.0), 0.95–0.70 (m, 2), 0.60 (q, 6, J = 8.0), 0.55-0.43 (m, 1); ¹³C NMR 201.2, 136.2, 133.6, 129.0, 125.9, 109.0, 106.2, 66.6, 53.9, 46.3, 35.3, 29.2, 28.0, 26.2, 25.3, 25.2, 25.0, 24.9, 23.1, 6.9 (3 C), 4.9 (3 C); HRMS (QTOF) calcd for C25H44NO2Si (MH+) 418.3136, found 418.3142.

X-ray Data Collection, Solution, and Refinement for 47b. All operations were performed on a modern, kappa-geometry X-ray diffractometer equipped with a CCD detector and graphite-monochromated Mo K α radiation. All diffractometer manipulations, including data collection, integration, scaling, and absorption corrections, were carried out using standard software.⁴⁴ Preliminary cell constants were obtained from three sets of 12 frames. Data collection was carried out at 120 K, using a frame time of 30 s and a detector distance of 60 mm. The optimized strategy used for data collection consisted of three phi and one omega scan sets, with 0.5° steps in phi or omega; completeness was 99.7%. A total of 2133 frames were collected. Final cell constants were obtained from the *xyz* centroids of 7440 reflections after integration.

From the systematic absences, the observed metric constants and intensity statistics, space group C2/c was chosen initially; subsequent solution and refinement confirmed the correctness of this choice. The structure was solved using direct methods⁴⁵ and refined by using fullmatrix-least-squares methods.⁴⁶ The asymmetric unit contains one complex (Z = 8; Z' = 1). All ordered non-hydrogen atoms were refined using anisotropic displacement parameters. After location of H atoms on electron-density difference maps, the H atoms were initially refined with soft restraints on the bond lengths and angles to regularize their geometry (C—H in the range 0.93–0.98 Å and U_{iso} (H) in the range 1.2–1.5 times U_{eq} of the parent atom), after which the positions were refined with riding constraints.⁴⁷ Minor ring disorder was observed at atom C(10); the two orientations were modeled as C(10)and C(101), and the occupancies were constrained to sum to 1.0. The major component occupancy (atom C(10), refined using an anisotropic displacement parameters) was 0.922(5); atom C(101) was refined using an isotropic displacement parameter. Atom H(1), attached to N(1) was refined using an isotropic displacement parameter. The final least-squares refinement converged to R_1 = 0.0434 ($I > 2\sigma(I)$, 3018 data) and wR₂ = 0.1117 (F^2 , 3333 data, 190 parameters). The final CIF is available as Supporting Material.

ASSOCIATED CONTENT

S Supporting Information

Copies of ¹H, ¹³C, and 2D NOESY NMR spectral data. Crystallographic data for compound **47b**. This material is available free of charge via the Internet at http://pubs.acs.org.

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Notes

The authors declare no competing financial interest.

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